

# **This student paper was written as an assignment in the graduate course**

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## Hydroxyl Radical

by

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### Abbreviations

HO: hydroxyl radical

2,3-DHBA: 2,3-dihydroxybenzoate

2,4-DHBA: 2,4-dihydroxybenzoate

2,5-DHBA: 2,5-dihydroxybenzoate

HPLC: high performance liquid chromatography

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### ***Abstract***

Hydroxyl radical ( $\text{HO}^\cdot$ ) is highly reactive species and is electrophilic in nature. Kinetics selectivity studies presented in this report support the hypothesis that  $\text{HO}^\cdot$  is the transient oxidant in the photo-Fenton reaction. Kinetics experiments for photo-Fenton reactions further indicate that  $\text{Fe(II)}$  and its oxalate, citrate, and phosphate complexes react with  $\text{H}_2\text{O}_2$  efficiently to produce  $\text{HO}^\cdot$  in aqueous solutions with pH ranging from 3 to 8. The present study has also investigated metal-binding to various  $\text{HO}^\cdot$  detectors, its effects on  $\text{HO}^\cdot$  measurement and scavenging, and the occurrence of site-specificity in the reaction of  $\text{HO}^\cdot$  with molecules.

## Introduction

Previous studies indicate that direct photolysis of nitrate and nitrite can be important sources of hydroxyl radical (HO·) in natural waters. In atmospheric water drops, photolysis of hydroxide complexes of Fe(III) is an important sources of HO·. In addition, abundant evidence exists that the Fe(II)-H<sub>2</sub>O<sub>2</sub> reaction is a major way for oxidation of Fe(II) and production of HO· in sunlit regions of surface waters and the sea [1].

## Chemical properties

The property of HO· has been developed by aqueous radiation chemistry. Evidence from table 1 shows that HO· is an oxidant. The HO· has a standard reduction potential of ~ 2.7 V in acidic solution and ~ 1.8 V in neutral solution [2].

**Table 1.** Properties of HO· in aqueous solution

Absorption maximum (nm)	~ 225
Extinction coefficient, $\epsilon$ (L mol <sup>-1</sup> cm <sup>-1</sup> )	540 (188 nm)
Charge	0
Radius (cm x 10 <sup>8</sup> )	2.2
Primary yield (molecules per 100 eV), pH 7	2.7
Diffusion coefficient (cm <sup>2</sup> s <sup>-1</sup> x 10 <sup>5</sup> )	2 ~ 2.3
Reduction potential (V)	1.77 ~ 1.91 (HO· + e <sup>-</sup> → OH <sup>-</sup> ) 2.59 ~ 2.74 (HO· + e <sup>-</sup> + H <sup>+</sup> → H <sub>2</sub> O)
pK <sub>a</sub>	11.9
$\Delta H$ (ionization) (kJ mol <sup>-1</sup> )	42
Electron affinity (eV)	1.83
$\Delta G_f^\circ$ (kJ mol <sup>-1</sup> )	13 ~ 35.7
$\Delta H_f^\circ$ (kJ mol <sup>-1</sup> )	-7 ~ -4
S <sup>o</sup> (J mol <sup>-1</sup> K <sup>-1</sup> )	96
$\Delta G$ (hydration)(kJ mol <sup>-1</sup> )	-21 ~ -10
$\Delta H$ (hydration)(kJ mol <sup>-1</sup> )	-42

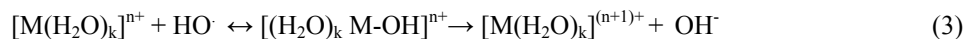
The simplest way to represents the reaction of HO· with ions is shown in equation 1 which contains a simple electron transfer. S means ion and n means the charge on the ion [2].



Also, intermediate adducts may be formed during the oxidation process of HO· and halide ions (X<sup>-</sup>), as shown in equation 2 [2].



In aqueous solution,  $\text{HO}\cdot$  may react with metal ion (M) such as  $\text{Ti}^+$ ,  $\text{Ag}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Fe}^{2+}$  and  $\text{Mn}^{2+}$ . Hence, its coordination number (n) will be increased, as shown in the following equation [2].



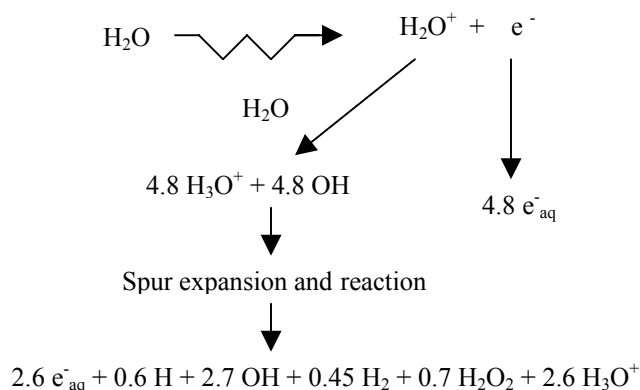
In strongly alkaline solution,  $\text{HO}\cdot$  rapidly converted to its conjugate base  $\text{O}^-$  with  $k = 1.2 \times 10^{10} \text{ L mol}^{-1} \text{ s}^{-1}$ , and  $\text{p}K_a(\text{HO}\cdot) = 11.9$  [2].



When  $\text{HO}\cdot$  reacts with organic molecules, it behaves as an electrophile. It adds to unsaturated bonds and abstracts H from C-H bonds readily. Finally, different products can be formed [2].

### ***Radiolysis of water***

There are two basic methods of generating  $\text{HO}\cdot$ . One is the radiolysis of water, and the other is Photo-Fenton reaction that relates to the iron-salt-dependent decomposition of  $\text{H}_2\text{O}_2$ . Scheme 1 represents the radiolysis of neural water at  $10^{-16}$  to  $10^{-7}$  s after irradiation. During above process, the initial radiolysis product is called spur. It expands through diffusion and reacts continuously. Also, one can regard  $10^{-7}$  s as the lifetime of  $\text{HO}\cdot$  which reacts with solutes at the diffusion-controlled rate in neural water [2].

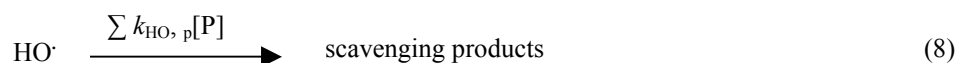
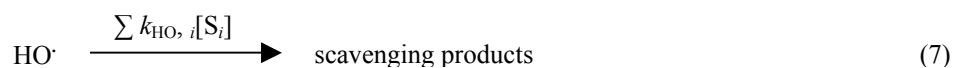


**Scheme 1.** Radiolysis of water

### ***Photo-Fenton reactions***

On the other hand, Fenton reaction occurs under acidic condition such as in atmospheric water droplets, freshwater and seawater. This reaction involves the generation of HO $\cdot$  from Fe(II) and H $_2$ O $_2$ . Previous studies indicate that Fe(II) complexes react with H $_2$ O $_2$  at neutral pH might not result in HO $\cdot$ . To determine Fenton reaction result in HO $\cdot$  occurs in acidic condition, kinetic approach was performed. Results showed Fe(II) complexes including Fe(II)-oxalates and Fe(II)-citrates can react with H $_2$ O $_2$  and produce HO $\cdot$  in aqueous solutions with pH ranging from 3 to 8 [1].

To further demonstrate HO $\cdot$  is the transient oxidant in the Fenton reaction, steady-state approach could be performed. One can irradiate aqueous system containing different probe compound until the reactive transients reach a steady state in which the transient's generation rates equal their decay rates. The model for Fenton reactions can be described by equations 5-8. In equation 5 and 6, Fe(III)L $_n$  represents the photoreactive complexes of Fe(III) in the system. In equation 7,  $\sum k_{HO, i}[S_i]$  is the pseudo-first-order rate constant (s $^{-1}$ ) for HO $\cdot$  scavenging by all constituents of the solution except the probe compound. In equation 8,  $\sum k_{HO, p}$  is the second-order rate constant (M $^{-1}$ s $^{-1}$ ) for reaction of HO $\cdot$  with the probe (P). As a result, the lifetime of the hydroxyl radical is determined by its reactions with the probe compounds and with scavengers (S $_i$ ) [1].



Fenton reactions can also be accelerated by the addition of reducing agents such as superoxide radical, ascorbate, and paraquat radical. Iron complex will become reduced forms and HO $\cdot$  will be decomposed. Both effects will generate the damaging species and cause more damage to the biological molecule [3].

### *Rate constants*

To demonstrate the rate constant of HO $\cdot$ , direct method and competition kinetics are used. In the direct method, the data are obtained by using pulse radiolysis and by measuring the decay of HO $\cdot$  directly or observing the formation of products, as shown in the following equation [2].



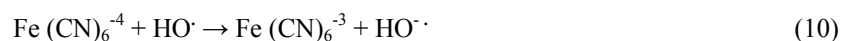
Actually, it's hard to measure the decay of HO $\cdot$  because of its weak ultraviolet absorption spectrum. Also, the formation of the product can only be observed when it has proper absorption spectrum. That's why competition kinetic method is generally used. There are three advantages of this method. 1) higher concentration of solutes can be performed. 2) eliminate the possibility of HO $\cdot$  reacting with itself. 3) shorten the timescale of observation and minimize the extent of continuous reactions of the products. Table 2 shows some established rate constants for HO $\cdot$ . Generally, for oxidation of many aquated metal cations, the rate constant of HO $\cdot$  is usually less than  $3 \times 10^8 \text{ L mol}^{-1}\text{s}^{-1}$  [2].

**Table 2.** Selected rate constants for HO $\cdot$

Reactant	$k$ (L mol $^{-1}$ s $^{-1}$ )
Bicarbonate ion	$8.5 \times 10^6$
Thiocyanate ion	$1.1 \times 10^{10}$
Carbonate ion	$3.9 \times 10^8$
Ferrocyanide ion	$1.05 \times 10^{10}$
Iodide ion	$1.1 \times 10^{10}$
Benzoate ion (BzO $^-$ )	$5.9 \times 10^9$
N, N-Dimethyl-4-nitrosoaniline (RNO)	$1.25 \times 10^{10}$
Ethanol	$1.9 \times 10^9$
Formate ion	$3.2 \times 10^9$
Methanol	$9.7 \times 10^8$
2-Methyl-2-propanol ( <i>tert</i> -BuOH)	$6.0 \times 10^8$
Nitrobenzene (NB)	$3.9 \times 10^9$
4-Nitrobenzoate ion (PNBA $^-$ )	$2.6 \times 10^9$
2-Propanol	$1.9 \times 10^9$
Thymine (5-MeU)	$6.4 \times 10^9$

Although there are several examples of HO $\cdot$  reacting with inorganic ions at the diffusion controlled rate, rate constants for oxidation many aquated metal cations usually less than  $3 \times 10^8$  L mol $^{-1}$  s $^{-1}$  [2].

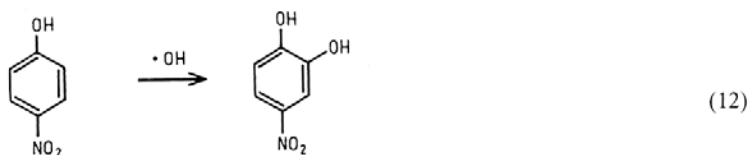
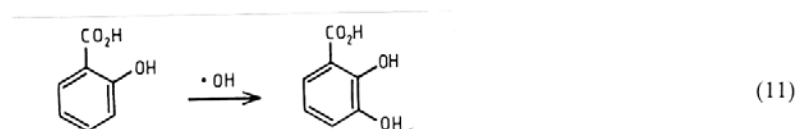
It is to be noted that the rate constant of HO $\cdot$  can still be measured by direct observation of the formation of the product Fe (CN) $_6^{3-}$  which is more stable compared to other reactants [4].



Moreover, ferrocyanide and thiocyanate ions have been used extensively as reference solutes in measuring rate constant of HO $\cdot$  by competition methods. For example, the values of rate constant for the reaction of hydroxyl radicals with ferrocyanide ions can be calculated from the psuedo-first order growth of the ferricyanide [5].

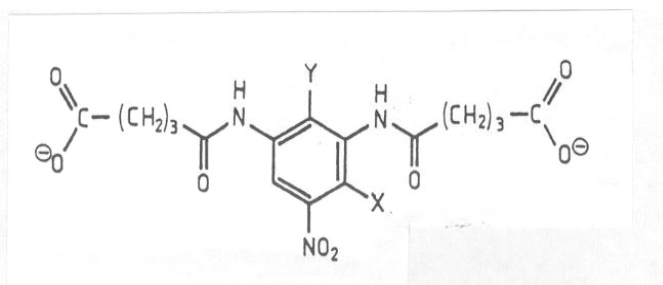
### *Detection*

Aromatic compounds are good detectors because they are hydroxylated to stable derivatives by HO $\cdot$  readily. In addition, the position of attack on the ring depends on the electron-donating or the withdrawing properties of the substituents. Shown in equation 11 and 12 are examples of the formation of hydroxylated aromatic products, salicylate and 4-nitrophenol [6].





As a result, one can use hydroxylation assay to detect  $\text{HO}^\bullet$  produced by various iron complexes in the Fenton reaction. This assay depends on the ability of these iron complexes to hydroxylate the aromatic substrate. As shown in Figure 1, the substrate (I) doesn't react with  $\text{HO}^\bullet$  quantitatively, it still can be used to compare  $\text{HO}^\bullet$  production in various solutions. On hydroxylation, the predominant products formed are the *ortho* and *para* hydroxy compounds (II) and (III). Both can be quantified by standard high performance liquid chromatography (HPLC) techniques [6].

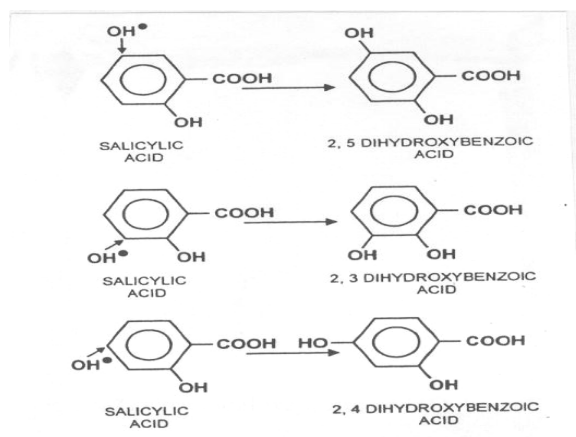


**Figure1.** Structure of N, N'-(5-nitro-1,3-phenylene) bisglutaramide (I), X=H, Y=H

N, N'-(4-hydroxy-5-nitro-1,3-phenylene)- bisglutaramide (II), X=OH, Y=H

N,N'-(2-hydroxy-5-nitro-1,3-phenylene) bisglutaramide (III), X=H, Y=OH

Spin trap labels is another method for  $\text{HO}^\bullet$  detection. The use of salicylate as a reporter is also based on the ability of  $\text{HO}^\bullet$  to attack aromatic compounds. Figure 2 shows the major products of  $\text{HO}^\bullet$  attack on salicylate are 2,3-dihydroxybenzoate (2,3-DHBA), 2,4-dihydroxybenzoate (2,4-DHBA) and 2,5-dihydroxybenzoate (2,5-DHBA). Different DHBAs can be separated and detected by HPLC.



**Figure2.** Chemical trapping of  $\text{HO}^\bullet$  by salicylate

Other biochemical assays for HO $\cdot$  detection include the production of formaldehyde from dimethylsulfoxide, the production of ethylene from methional, oxidation of formate to carbon dioxide, bleaching of p-nitrodimethylaniline [6].

One should keep in mind is that when two or more HO $\cdot$  detectors or scavengers are present in an assay system they will compete for available HO $\cdot$ . For example, when phenylalanine competed with salicylate, DHBA production was consistently lower than calculated for random competition. This suggested that other factors may be involved in the competition for HO $\cdot$  [5].

## References

1. Zepp RG, Fauset BC, Hoigne J. (1992) Hydroxyl radical formation in aqueous reaction (pH 3-8) of iron(II) with hydrogen peroxide: The photo-Fenton reaction. *Environ Sci Technol.* **26**:313-319.
2. George VB. (1988) Critical review of rate constants for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals (HO<sup>•</sup>/O<sup>•</sup>) in aqueous solution. *J. Phys. Chem. Reference Data* **17**:513-886.
3. Aruoma OI, Halliwell B, Gajewski E, Dizdaroglu. (1989) Deoxyribose assay for detecting hydroxyl radicals. *J Biol Chem.* **264**:57-66.
4. Allen JE, Anita SS. (1983) Rate constants for reactions of hydroxyl radicals as a function of temperature. *Radiat Phys Chem.* **24**:229-231.
5. Gelvan D, Moreno V, Gassmann, Hegenauer J, Saltman P. (1992) Metal-ion-directed site-specificity of hydroxyl radical detection. *Biochim Biophys Acta.* **1116**:183-191.
6. Singh S, Hider RC. (1988) Colorimetric detection of the hydroxyl radical: comparison of the hydroxyl-radical-generating ability of various iron complexes. *Anal Biochem.* **171**:47-54.
7. Powell SR. (1994) Commentary salicylate trapping of HO<sup>•</sup> as a tool for studying post-ischemic oxidative injury in the isolated rat heart. *Free Rad Res.* **21**:355-370.