

This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2005)

offered by the

Free Radical and Radiation Biology Program

B-180 Med Labs

The University of Iowa

Iowa City, IA 52242-1181

Spring 2005 Term

Instructors:

GARRY R. BUETTNER, Ph.D.

LARRY W. OBERLEY, Ph.D.

with guest lectures from:

Drs. Freya Q. Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Glutathione disulfide reductase : reducing oxidative stress



Disha Dayal

Free Radical and Radiation Biology Program

University of Iowa

Iowa City, IA-52242-1181

Overview



1. Introduction

1.1 About the enzyme

1.2 Reaction catalyzed

2. Protein structure

3. Mechanism of reaction

4. Studying GR

5. Enzyme regulation and stability

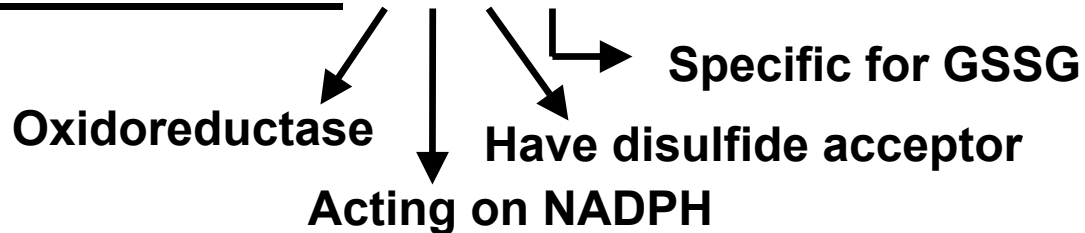
6. Summary

About the enzyme



Alternative names: Glutathione reductase, NADPH-glutathione reductase, GSH reductase, GSSG reductase, usually abbreviated as GR.

EC number*: 1.6.4.2



Isolated from: Yeast (*Saccharomyces cerevisiae*), spinach, erythrocytes *etc.*

Active site: Cys58-Cys63-His467

***Dimer molecular weight: 105 kDa**

***Isoelectric point: 6.46**

Catalytic efficiency: 2 M⁻¹s⁻¹

$\epsilon_{461}: 11.7 \times 10^{-3} \text{ M}^{-1} \text{ cm}^{-1}$

Turnover for GSSG: 9900 min⁻¹

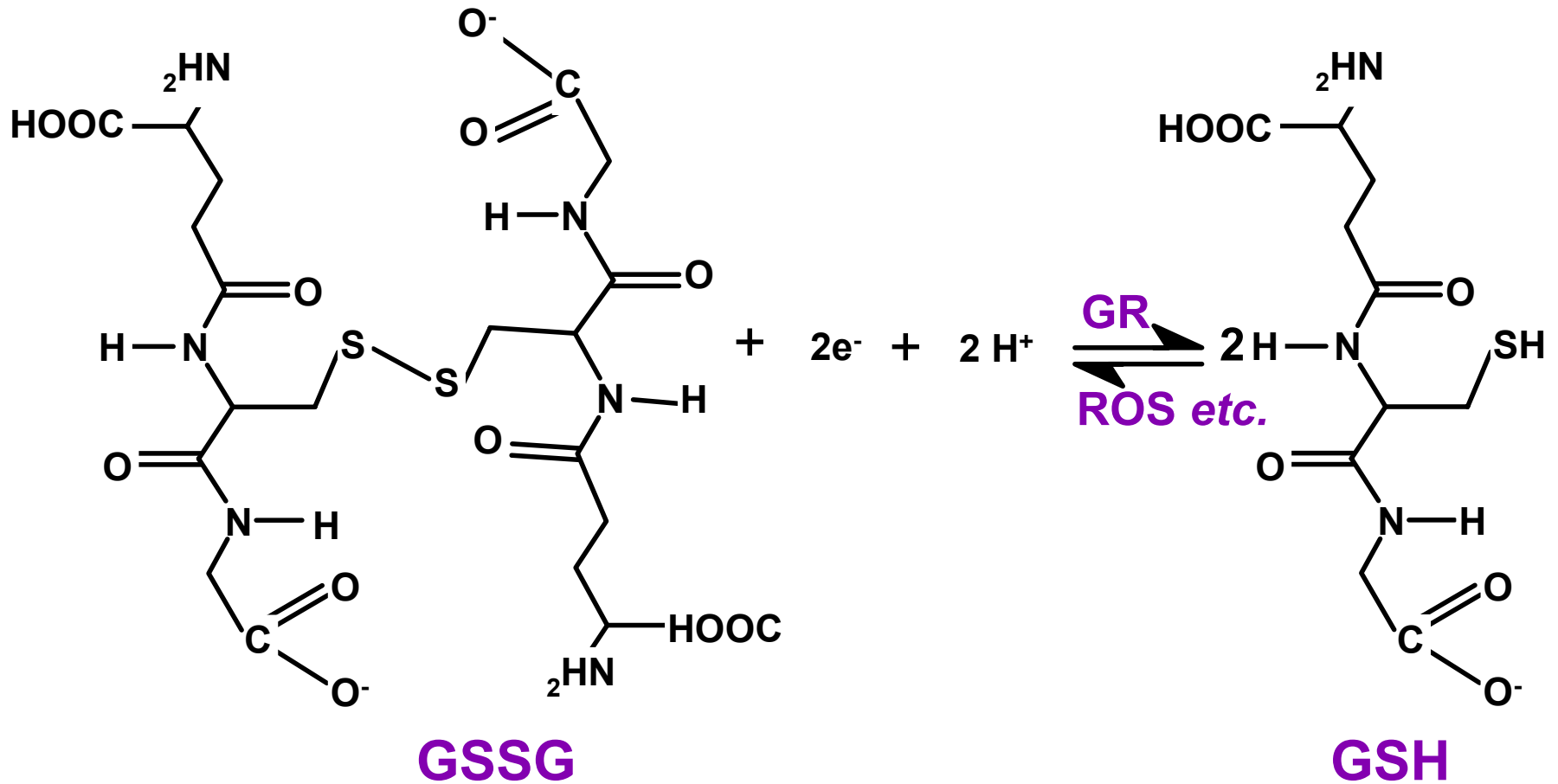
Prosthetic group: FAD

Cofactor: NADPH

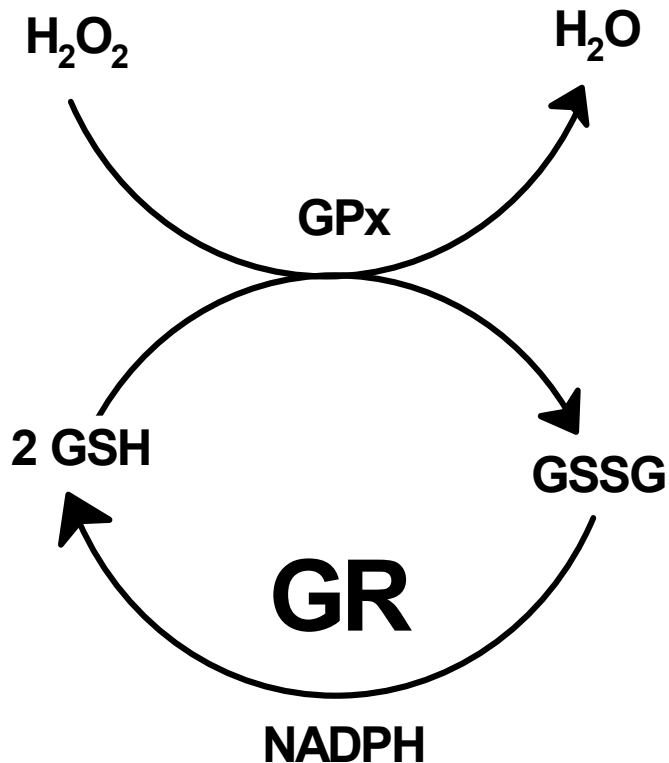
Mol Biochem Parasitol (2000) **107**:169–179; * *Eur J Biochem.*(1979) **98**:487-99.



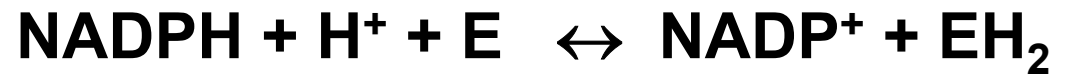
Reaction catalyzed



Reaction equation



Step 1:

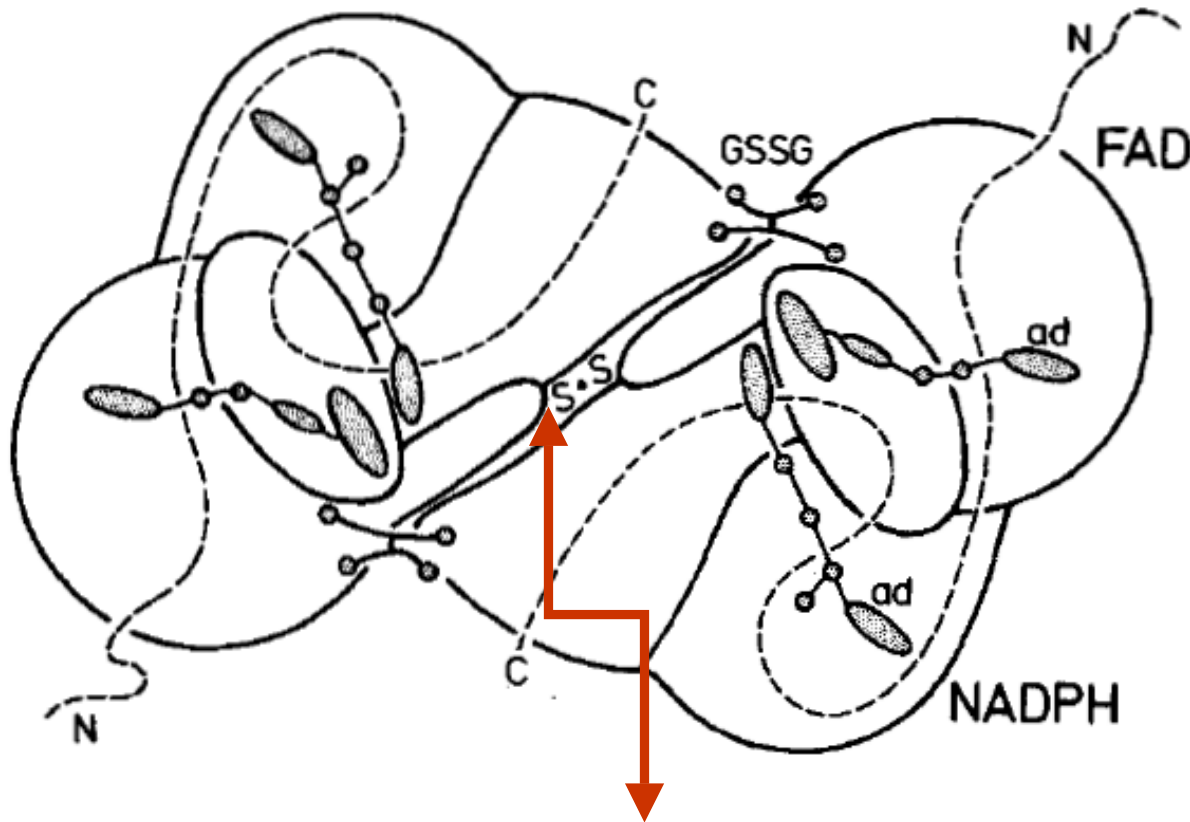


Step 2:



Where E = enzyme

GR has two equivalent subunits:
Catalytic center lies between these
two subunits

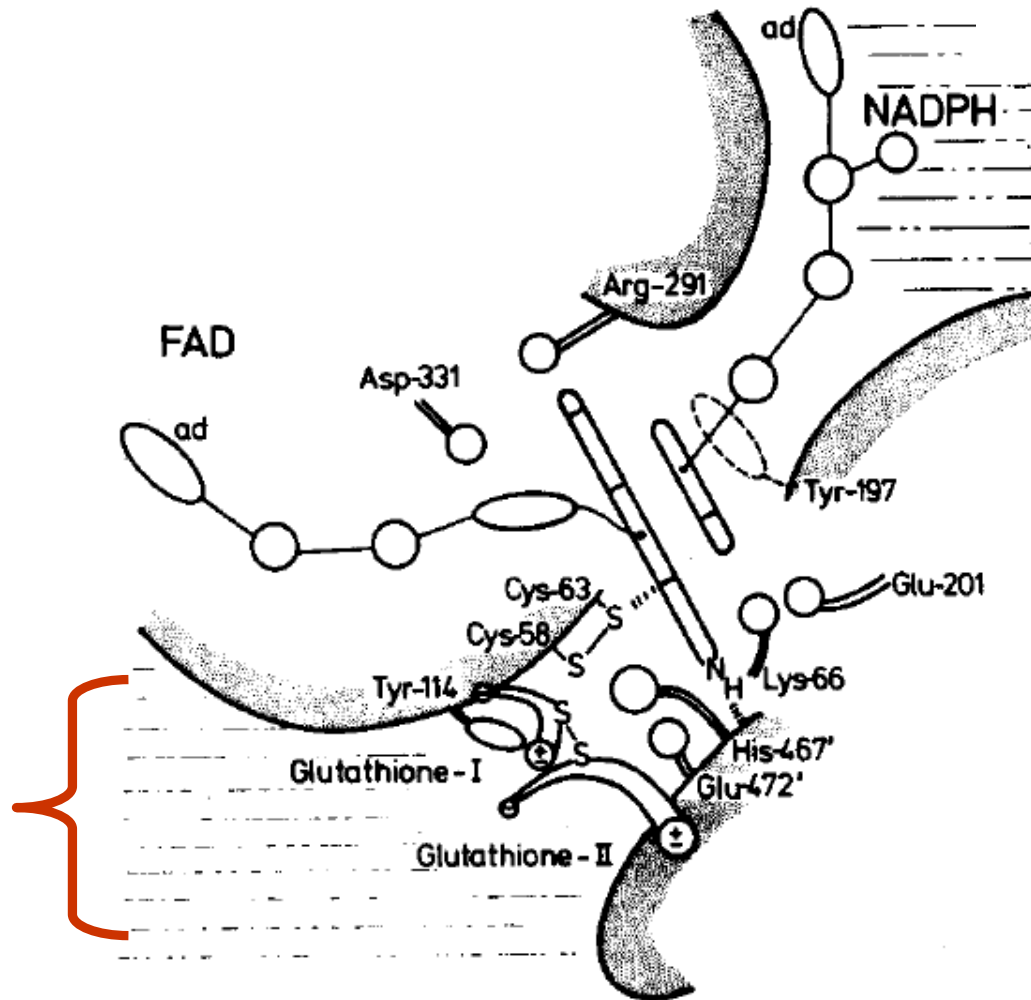


Catalytic center

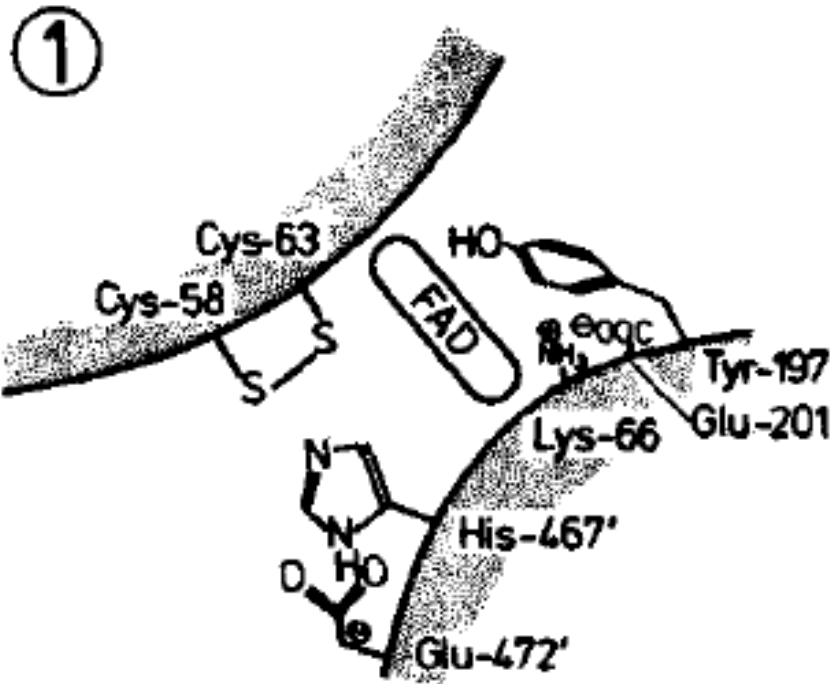
Catalytic center is divided into two parts: one binds NADPH and the other binds GSSG



Substrate
GSSG
represented as
GSH I:GSH II



Mechanism step 1

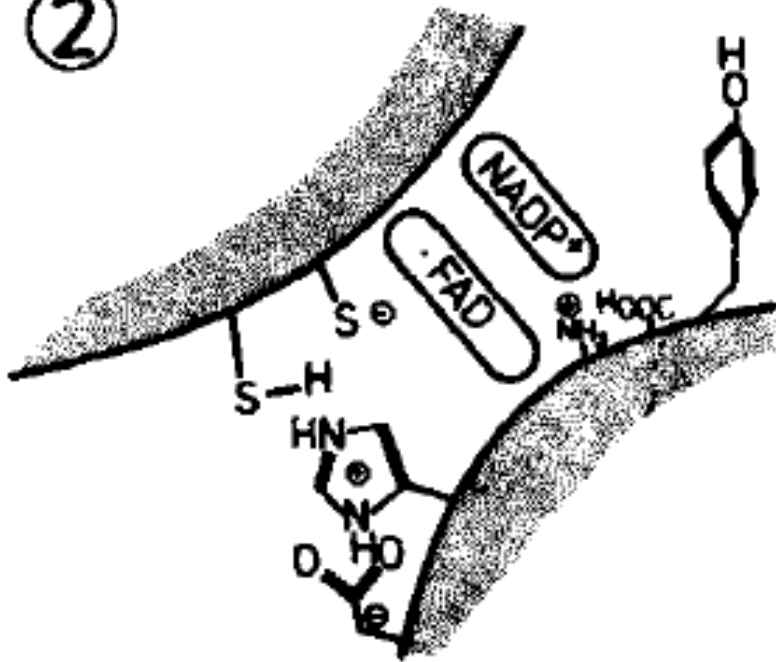


- Oxidized form of the enzyme.
- The -N of His 467 bonds with -OH of Glu 472.

Mechanism step 2

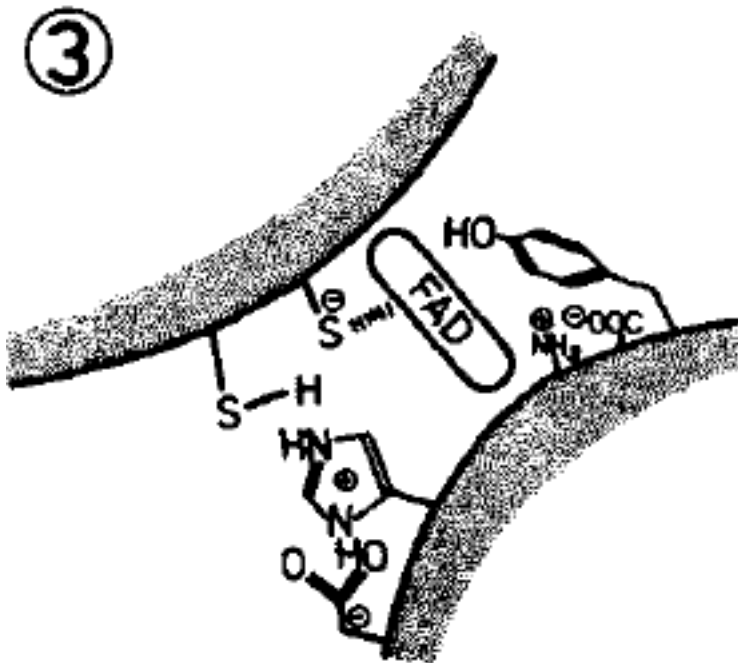


②



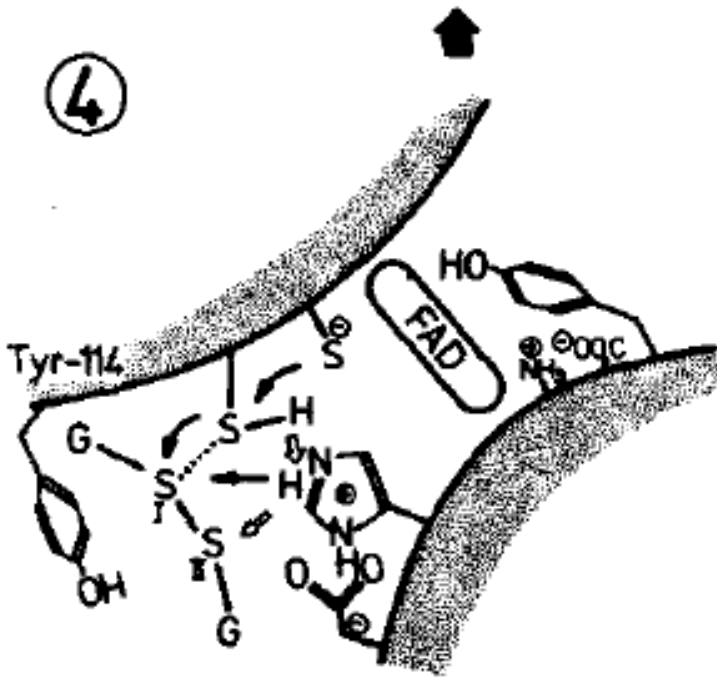
- NADPH reduces the enzyme to EH₂.
- On binding NADPH, Tyr 197 in the NADPH binding pocket moves away such that flavin of FAD can interact with nicotinamide of NADPH.

Mechanism step 3



- Sulfur of Cys-63 moves closer to FAD forming the charge transfer complex.

Mechanism step 4

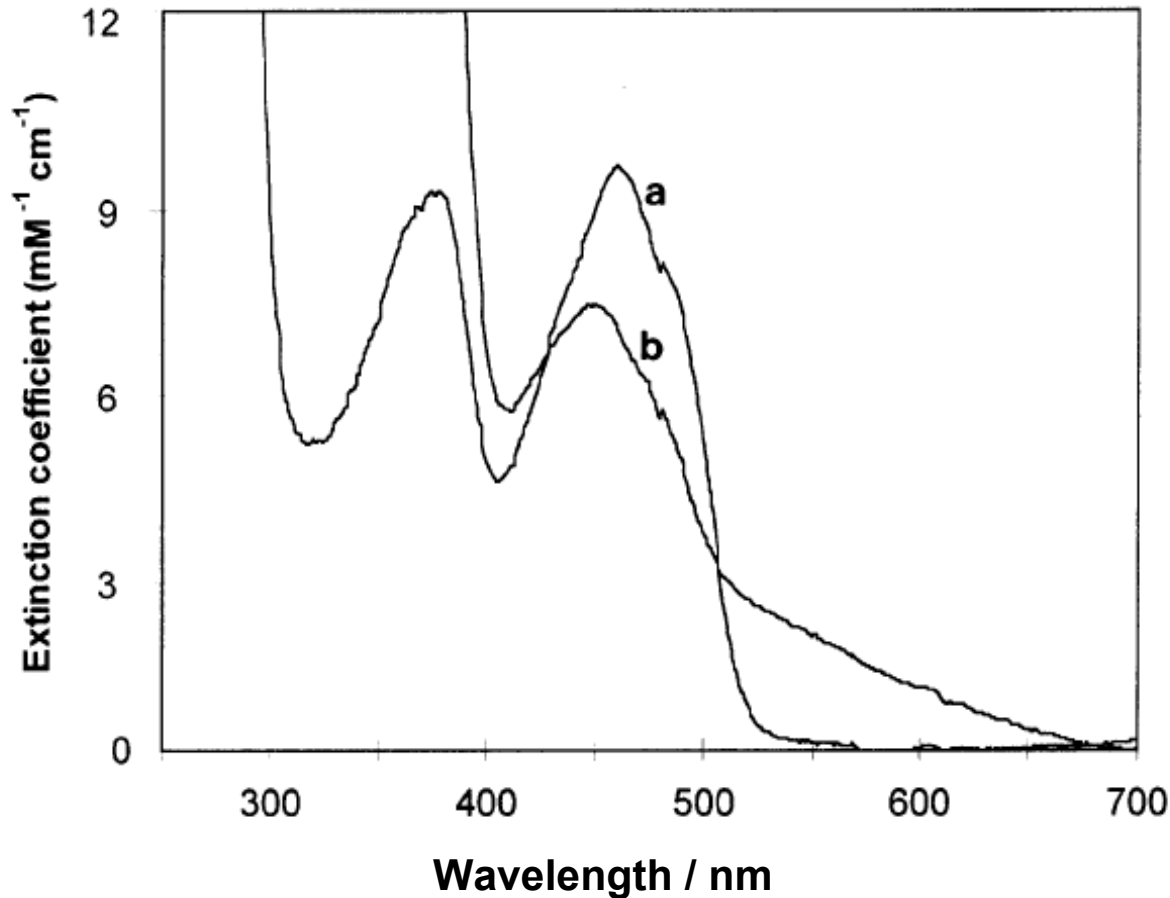


- GSSG binds to reduced enzyme EH_2
- Glutathione I and Cys-58 form a mixed disulfide releasing glutathione II
- In the last step, glutathione I is released

Studying GR:



(1) Absorption spectra of GR



a: reduced form
b: oxidized form

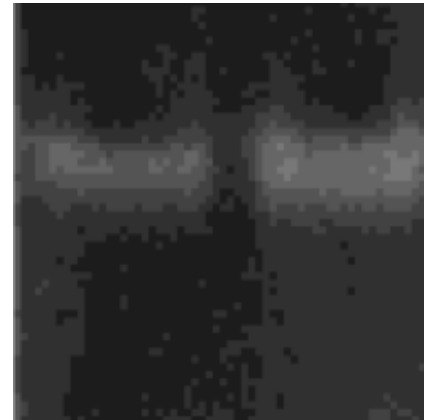
**Enzyme
purity/activity
can be tested by
checking its ϵ_{461}**

(2) Activity assay



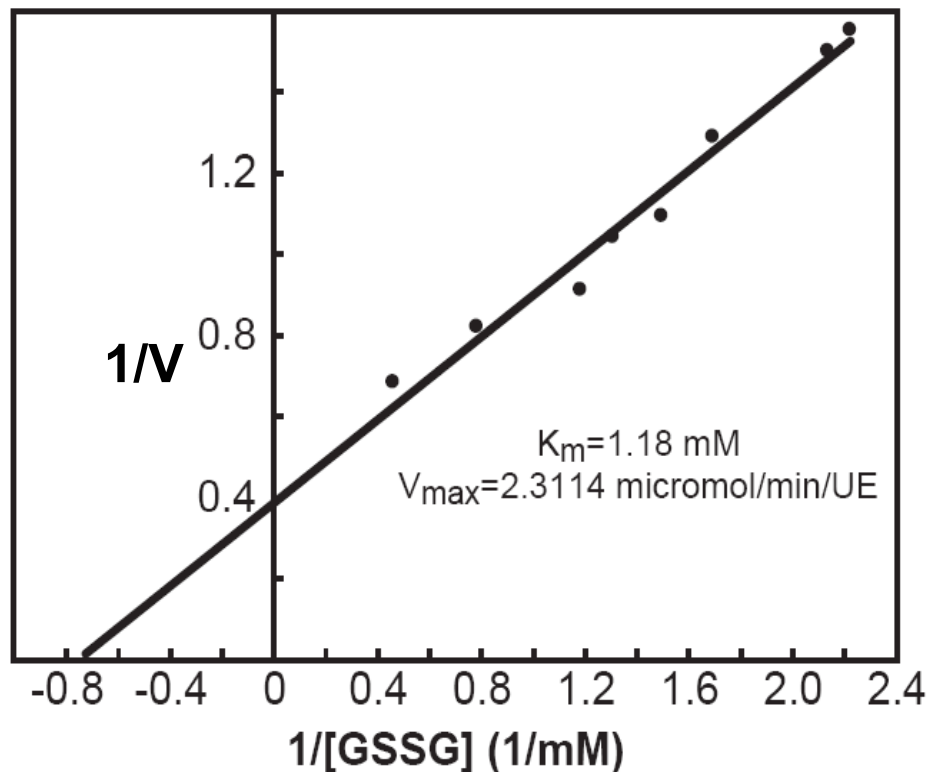
Activity can be followed
by native PAGE using
MTT/PMS* negative
staining.

0.8 U 0.32 U



***MTT**: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide;
PMS: phenazine methosulfate

(3) Lineweaver-Burk plots determined polarigraphically help in V_{max} and K_m determination



Average $V_{max} = 2.30 \pm 0.01$

$\mu\text{mol}/\text{min}/\text{UE}$ at

(25°C, pH = 8.0)

Average $K_m = 1.17 \pm 0.01$ mM

Enzyme regulation



- **The gene encoding GR is mostly post-transcriptionally regulated.**
- **GR protein levels are dictated by the metabolic needs of the cell and oxidative stress.**
- **Activity of the enzyme can also vary with temperature and pH.**

Enzyme stability



➤ **Prolonged storage (6 -12 months) of the enzyme at -20°C or -80°C reduces its activity by ~ 2%** *Clin chem.* (2000) **46:** 566-567.

➤ **On standing at room temperature, GR activity decreases. However, dilution of the enzymes seems to inhibit enzyme decay** *Clin chem.* (1976) **22:** 1005-1008.

Summary 1



- **GR is indirectly involved in combating oxidative stress by replenishing the reducing equivalents of the cell, namely, GSH. Thus it is a secondary antioxidant enzyme.**

- **It is widely studied and most of its physical and chemical properties are well-known.**

Summary 2



- It holds an important place at all levels of life plants, yeast and humans. Thus, there is significant homology in the GR genetic code at all levels.
- Further studies need to be done to assess its regulation by competitive and non-competitive inhibition in the cell which might reduce its turnover.
- Loss of GR activity can prove to be detrimental to the cell.
- Thus GR inhibition forms a potential target for anti-cancer therapies.