

This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2005)

offered by the

Free Radical and Radiation Biology Program

B-180 Med Labs

The University of Iowa

Iowa City, IA 52242-1181

Spring 2005 Term

Instructors:

GARRY R. BUETTNER, Ph.D.

LARRY W. OBERLEY, Ph.D.

with guest lectures from:

Drs. Freya Q. Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Nitroxides as Antioxidants, beyond Spin Probes

By

Changbin Du

Free Radical and Radiation Biology Program

Department of Radiation Oncology

The University of Iowa, Iowa city, IA, 52242-1181

For 77:222, Spring 2005

24 Feb 2005

Abbreviations:

CHD, spiro [cyclohexane- 1,2'-doxyl] [spiro[cyclohexane- 1,2'-(4',4'-dimethyl-oxazolidine-3'-oxyl)]. **DMPO**, 5,5-dimethyl-1-pyrroline-N-oxide. **EPR**, electron paramagnetic resonance. **ESR**, electrons spin resonance. **MRI**, magnetic resonance imaging. **OXANO**: 2-ethyl-2,5,5-trimethyl-3-oxazolidinoxyl. **O₂^{•-}**, superoxide. **OXANOH**: 2-ethyl-2,5,5-trimethyl-3-oxazolidine. **•OOH**, protonated superoxide. **PNA**, polynitroxyl-albumin. **TPO**, 2,2,6,6-tetramethyl-piperidinoxyl. **TPL**, 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-oxyl. **TPH**, 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-hydroxyl.

Outline	Page
Abstract.....	2
Introduction	3
The chemical properties of nitroxides.....	3
Nitroxides as antioxidants and mechanism.....	5
Summary.....	10
References.....	10

Abstract

Stable nitroxide free radicals have been employed to probe various biophysical and biochemical processes involving oxidative stress. Previous studies demonstrated that nitroxides at non-toxic concentrations are effective as *in vitro* and *in vivo* antioxidants. Therefore, studying the mechanism underlying their antioxidant activity will help to improve therapeutic strategies by nitroxides. The capability of nitroxides to remove intracellular superoxides, to terminate radical-chain reactions, to oxidizes deleterious metal ions and prevent Fenton reaction may contribute their distinguished functions as antioxidants.

Introduction

Nitroxide stable free radicals or nitroxyl spin labels as they may be called, are chemically stable organic compounds that have an unpaired electron resulting in paramagnetic properties. Because of this, they have been used as imaging agents for electron spin resonance (ESR) studies and as contrast agents for magnetic resonance imaging (MRI) in animals [2]. An increasing number of studies demonstrated that nitroxides possess antioxidant functions, which suggested their important therapeutic potential in a wide range of diseases. In this paper, some chemical properties of nitroxides and their antioxidant functions and the mechanism behind that will be discussed.

The chemical properties of nitroxides

Nitroxides generally consist of a six-member ring piperidine derivative (such as TEMPO and TPO) or a five-member ring pyrroxamide derivative (such as ONANO) (Figure 1). The unpaired electron of the nitroxide moiety can be detected by EPR spectroscopy (Figure 2). Nitroxides are usually quite stable in aqueous solution, especially in alkali solutions [5]. The persistent nitroxide can still maintain signal intensity even at 250°C. Steric blocking methyl groups or alkyl groups may contribute to the kinetic stability [5]. Despite that rather immobilized electron in nitroxides, they can still undergo various reactions. Morris *et al* compared the reduction rates of five-membered pyrrolidine and pyrroline, and six-membered piperidine nitroxides. Their results showed an increased rate of reduction of six-membered nitroxides compared with those of the five-membered nitroxides [6]. The difference in the accessibility of their nitroxide group may contribute to this effect. A double bond in the five-membered nitroxyls increases the reduction rate

[6]. Results from Fuchs *et al* showed that the rate of reduction of nitroxides in skin varies considerably with nitroxide ring structure and substitution. The order of nitroxide stability in isolated human keratinocytes and human skin is imidazoline>pyrrolidine>piperidine>oxazolidine [7]. Cationic nitroxides are most unstable likely due to transmembrane electron shuttle or internalization [7].

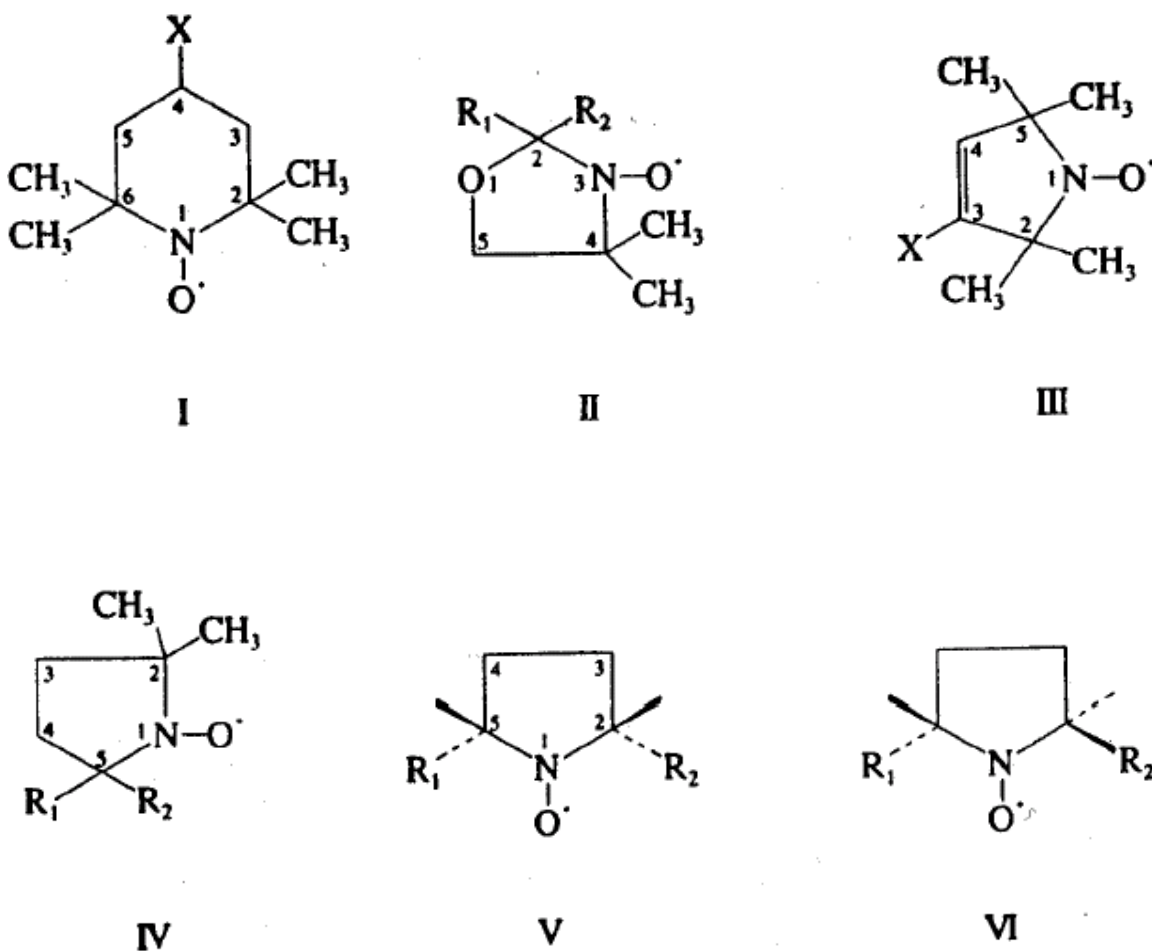


Figure 1. Structural classes of nitroxides: (I) piperidine; (II) oxazolidine; (III) pyrroline; (IV-VI) pyrrolidine. From [1].

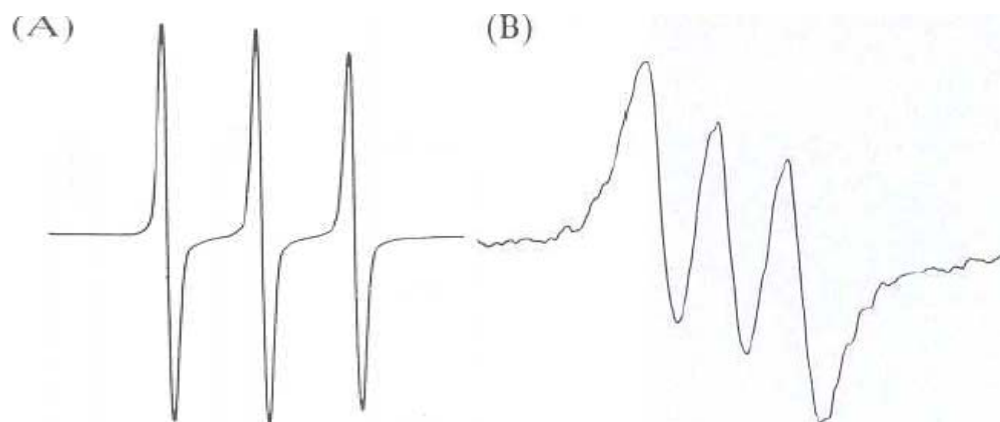
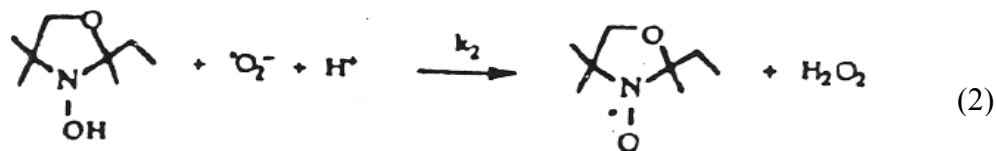
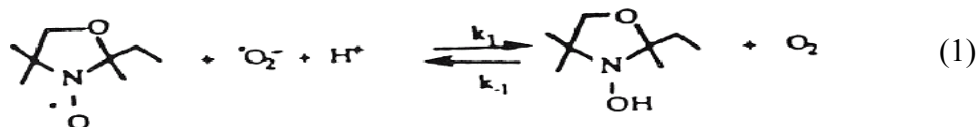


Figure 2: A, First derivative electron paramagnetic resonance spectrum of 5 mM TEMPOL in MEM in an EPR flat cell at ambient temperature. B, EPR spectrum of TEMPOL in the presence of broadening agent (100 mM $\text{K}_3\text{Cr}(\text{C}_2\text{O}_4)_3$) in MEM [4].

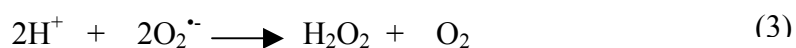
Nitroxides as antioxidants and mechanism

Remove $\text{O}_2^{\cdot-}$, nitroxides acts as scavengers of $\text{O}_2^{\cdot-}$ or as SOD mimics?

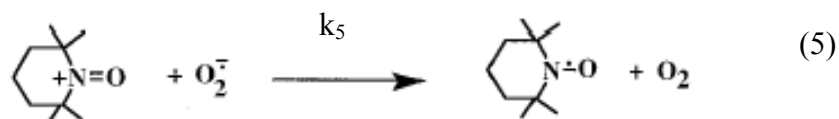
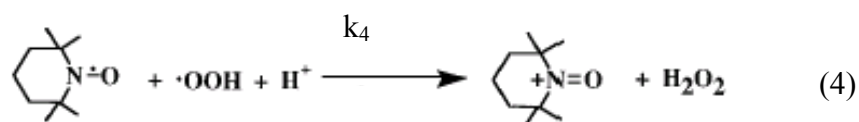
Nitroxides were found to be reduced to the corresponding hydroxylamine by $\text{O}_2^{\cdot-}$. This finding originally stemmed from the observation of $\text{O}_2^{\cdot-}$ -induced spin-loss of OH spin-adduct of 5,5-dimethyl-1-pyrroline-N-oxide (DMPO). The ESR signals of persistent nitroxide spin-adducts such as DMPO-OH rapidly decayed to a lower steady state when exposed to a continuous $\text{O}_2^{\cdot-}$ -flux [8]. Oxazolidine nitroxides are supposed to dismutate $\text{O}_2^{\cdot-}$ through the following reaction pathways.



Superoxide reacts with OXANO through one electron reduction to yield diamagnetic (“ESR-silent”) hydroxylamine OXANOH (Reaction 1). OXANOH and $\text{O}_2^{\cdot-}$ reacts to yield OXANO (Reaction 2) [9]. When OXANO reacts with $\text{O}_2^{\cdot-}$, rate constants k_1 and k_2 are 1.3×10^2 and $6.7 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$ respectively at pH 7.8 [9]. When TEMPO or TEMPOL react with $\text{O}_2^{\cdot-}$, rate constants at pH 7.8 range from $7 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$ to $1.2 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$ [8]. The overall reaction is the following:



Piperidine derivatives such as TPO and TPL are readily oxidized by $\cdot\text{OOH}$ (protonated superoxide) to yield oxoammonium cation (Reaction 4), which in turn oxidizes another $\text{O}_2^{\cdot-}$ to molecular oxygen (Reaction 5) [10]. The reaction rate constants k_4 were $7.4 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$ and $2.0 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$ for TPO and TPL, respectively [10]. Reaction 4 is the rate-limiting step in the catalytic dismutation of $\text{O}_2^{\cdot-}$ by nitroxides, because the second step (Reaction 5) is much faster having $k_5 = 1.5 \times 10^{10} \text{ M}^{-1}\text{s}^{-1}$ [10].



The net process of reaction 4 & 5 is reaction 3, in which $\text{O}_2^{\cdot-}$ induced depletion and regeneration of the nitroxides, and $\text{O}_2^{\cdot-}$ is removed without affecting nitroxides concentration. The above description supports the idea that nitroxides acts as SOD Mimics.

In the presence of reducing agents such as NADH and NADPH, transient oxoammonium cation was reduced through a two-electron reduction to its corresponding hydroxylamin (Reaction 6) [10].



If only 1 mM NADH was added to the mix of $\text{O}_2^{\cdot-}$, and TPO, EPR signal decreased rapidly (Figure 3) [10]. From those results, nitroxides should be regarded as scavengers rather than SOD mimics.

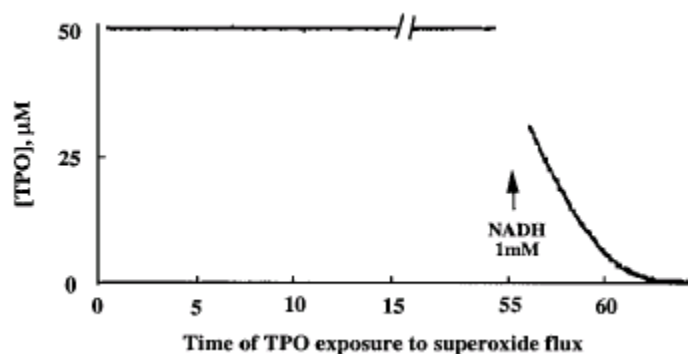
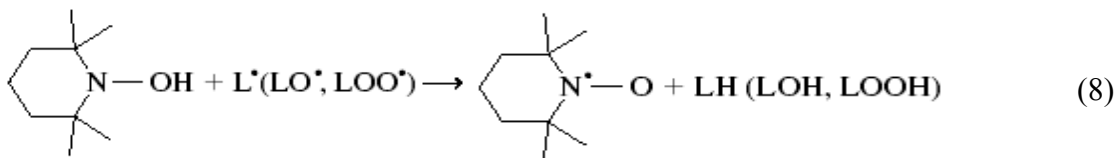
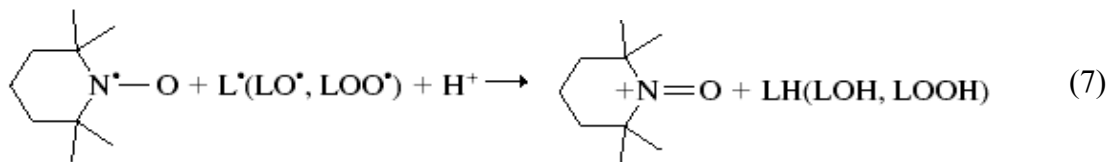


Figure 3. Invariance of the SOD mimic TPO subjected to superoxide flux over time. A constant flux of ~ 16 mM/min O_2 was generated at $23 \pm 2^\circ C$ by 5 mM HX and 0.08 unit/mL XO in 50 mM phosphate buffer, pH 7, containing 50 mM TPO, 50 mM DTPA, and 260 units/mL catalase. The reaction mixture was inserted in a gas-permeable capillary, placed inside the EPR cavity, and flushed with air, whereas the intensity of TPO EPR signal was continuously monitored. Upon the addition of 1 mM NADH to the reaction mixture (*arrow*), the EPR signal of TPO decayed [10].

Terminating radical-chain reactions

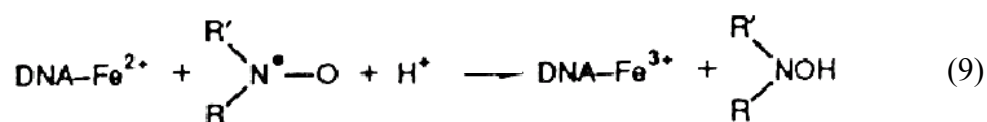
Both nitroxide and hydroxylamine can reduce carbon-centered and oxygen-centered radicals and break the radical chain reaction (Reaction 7 & 8) [13].



The high efficacy of both the nitroxide and its respective reduced form seem to result from the catalytic mode of their action. While switching back and forth between themselves (Reaction 1 & 2), stable nitroxides can terminate chain reaction of deleterious radicals with higher efficacy [13].

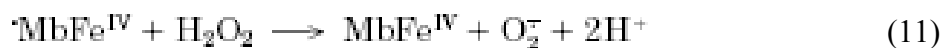
Preempt the Fenton reaction

Previous studies have shown that stable nitroxides are effective inhibitors of H₂O₂ - induced damage in mammalian cells [11]. Tissue injury can enhance iron mobilization and to increase the iron pool and free radical formation [13]. Mitchell found that, in the presence of CHD, DNA-Fe(II) reacted preferentially with CHD (reaction 9), which blocked the metal-catalyzed reduction of H₂O₂ to [•]OH which is highly reactive and can cause damage to DNA [12].



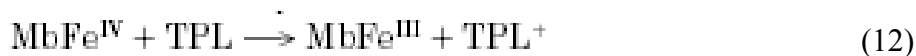
Detoxification of hypervalent toxic metal species

Oxymyoglobin (oxyMbFe(II)), the major heme protein in muscle tissue, can undergo oxidation and yield O₂^{•-} and MbFe(III) [15]. H₂O₂ and MbFe(III) has been shown to produce a two-electron oxidation product of myoglobin by a heterolytic cleavage of the O-O bond of the peroxide coordinated to the heme (Reaction 10). The oxoferryl species are chemically reactive and can initiate free radical-mediated reactions that could result in biologic damage [14].



Besides this, the ferryl moiety can oxidize another H₂O₂ molecule to O₂^{•-} (Reaction 11).

MbFe(IV) can oxidize critical targets of cells.



Nitroxides can reduce and detoxify MbFe(IV), yielding the oxoammonium cation TPL^{·+}, which can oxidize another H₂O₂ through Reaction 13 or generate O₂ by oxidizing O₂^{·-} through reaction 14 [14].

Summary

Nitroxides are chemically stable free radicals and can be detected by EPR spectra. The protection of oxidative damage by non-toxic levels of nitroxides have several plausible chemical explanations: 1) Remove superoxides. 2) Termination of free radical chain reactions. 3) Preempt the Fenton reaction. 4) Detoxification of hypervalent toxic metal species.

Reference

1. Kocherginsky N, Swartz HM. (1995) Terminology, classification and distribution of the nitroxides in cells. In: *Nitroxide Spin Labels Reaction in Biology and Chemistry*. Boca Raton: CRC Press; Chapter2: pp 15-25.
2. Nakagawa K, Ishida SI, Yokoyama H, Mori N, Niwa SI, and Tsuchihashi N. (1994) Rapid free radical reduction in the perfused rat liver. *Free Rad.Res.***21**:169-176.
3. Chyla AK, Gwozdziński K, Kochman A, Stolarska A, Jozwiak Z. (2003) Effects of pyrroline and pyrrolidine nitroxides on lipidperoxidation in heart tissue of rats treated with doxorubicin. *Cell & Mol Biol Lett.* **8**: 179-183.
4. Reddan JR, Sevilla MD, Giblin FJ, Padgaonkar V, Dziedzic DC, Leverenz V, Misra IC, Peters, JL. (1993) The superoxide dismutase mimic TEMPOL protects

- cultured rabbit lens epithelial cells from hydrogen peroxide insult. *Exp Eye Res.* **56**: 543-554.
5. Kocherginsky N, Swartz HM. (1995) Chemical reactivity of nitroxides. In: *Nitroxide Spin Labels Reaction in Biology and Chemistry*. Boca Raton: CRC Press; Chapter3: pp 25-65.
 6. Morris S, Sosnovsky G, Hui B, Huber CO, Rao NU, Swartz HM. (1991) Chemical and electrochemical reduction rates of cyclic nitroxides (nitroxyls). *J Pharm Sci.* **80**:149-52.
 7. Fuchs J, Freisleben HJ, Podda M, Zimmer G, Milbradt R, Packer L. (1993) Nitroxide radical biostability in skin. *Free Radic Biol Med.* **15**:415-423.
 8. Samuni A, Krishna CM, Mitchell JB, Collins CR, Russo A. (1990) Superoxide reaction with nitroxides. *Free Rad Res Comms* **9** (3-6): 241-249.
 9. Samuni A, Krishna CM, Riesz P, Finkelstein F and Russo A. (1988) A novel metal-free low molecular weight superoxide dismutase mimics. *J Biol Chem.* **263**: 17921-17924.
 10. Krishna MC, Russo A, Mitchell JB, Goldstein S, Dafni H, Samuni A. (1996) Do nitroxide antioxidants act as scavengers of $O_2^{\cdot-}$ or as SOD mimics? *J Biol Chem.* **271**:26026-31.
 11. S.M. Hahn, J.B. Mitchell, E. Shacter. (1997) Tempol inhibits neutrophil and hydrogen peroxide-mediated DNA damage. *Free Radic Biol Med.* **23**: 879-884.
 12. Mitchell JB, Samuni A, Krishna MC, DeGraff WG, Ahn MS, Samuni U, Russo A. (1990) Biologically active metal-independent superoxide dismutase mimics. *Biochemistry* **29**:2802–2807.
 13. Zhang R, Shohami E, Beit-Yannai E, Bass R, Trembovler V, Samuni A. (1998) Mechanism of brain protection by nitroxide radicals in experimental model of closed-head injury. *Free Radic Biol Med.* **24**:332-340.
 14. Krishna, M. C.; Samuni, A.; Taira, J.; Goldstein, S.; Mitchell, J. B.; Russo, A. (1996) Stimulation by nitroxides of catalase-like activity of heme proteins. Kinetics and mechanism. *J. Biol. Chem.* **271**:26018–26025.
 15. Tajima, G., and Shikama, K. (1987) Autoxidation of oxymyoglobin. An overall stoichiometry including subsequent side reactions. *J. Biol. Chem.* **262**, 12603–12606.