# ON THE REACTION OF SUPEROXIDE WITH DMPO/ OOH

# GARRY R. BUETTNER

ESR Center, EMRB 58, The University of Iowa, Iowa City, IA 52242 USA

A kinetic model has been used to estimate the rate constant for the reaction of superoxide ( $O_2^-$ /'OOH) with the superoxide spin adduct of 5,5-dimethylpyrroline-N-oxide, DMPO/'OOH. This rate constant is estimated to be 4.9 ( $\pm$  2.2)  $\times$  10<sup>6</sup> M<sup>-1</sup> s<sup>-1</sup>, pH 7.4 and 25°C.

KEY WORDS: Spin trapping, 5,5-dimethylpyrroline-N-oxide, superoxide, free radical.

ABBREVIATIONS: DETAPAC, diethylenetriaminepentaacetic acid; DMPO, 5,5-dimethylpyrroline-Noxide; 3-CP, 3-carboxy-proxyl; OXANO, 2-ethyl-2,5,5-trimethyl-3-oxazolidinoxyl; OXANOH, 2-ethyl-1-hydroxy-2,5,5-trimethyl-3-oxazolidine; X.O., xanthine oxidase.

### INTRODUCTION

Superoxide\* reacts slowly with DMPO at neutral pH ( $k^{obs} = 30 \, M^{-1} \, s^{-1}$  at pH 7.4<sup>1</sup>) producing a spin adduct, DMPO/OOH, that decays by a first-order process, and is relatively short-lived ( $t_{1/2} = 50 \, s$  at pH 7.4, 25°C²). It has recently been shown that the reaction of  $O_2^-$  with DMPO/OH and DMPO/CH<sub>3</sub> may be a significant process and should be considered when interpreting spin trapping data.<sup>3,4</sup> In general, it was found that 5-membered ring nitroxides react with superoxide to produce diamagnetic products. Thus, it is reasonable to suspect that superoxide will react with DMPO/OOH. However, the short lifetime of DMPO/OOH precludes a simple direct determination of the rate constant for the reaction:

# $O_{7}^{-}$ + DMPO/OOH $\rightarrow$ products.

However, by: 1) determining the rate of production of superoxide in a superoxidegenerating system; 2) determining the steady-state concentration of DMPO/OOH; and 3) using an appropriate kinetic model, I have estimated the rate constant for this reaction.

# MATERIALS AND METHODS

Xanthine oxidase, hypoxanthine, cytochrome c, 3-carboxy-proxyl, and DMPO were from Sigma. DMPO was purified with charcoal<sup>2</sup> and its concentration determined using  $\varepsilon_{228} = 7.8 \times 10^3 \, \mathrm{M}^{-1} \, \mathrm{cm}^{-1}$  (G.R. Buettner, unpublished). Adventitious metals were removed from the buffer with chelating resin (sodium form, dry mesh 50–100, from Sigma, St. Louis, MO). In the demetalled buffer, the loss of ascorbate was 0.3% or less in the standard 15 minute test, indicating effective removal of catalytic metals.

<sup>\*</sup>In this paper, I use superoxide (or  $O_2^-$ ) to represent the equilibrium mixture of  $O_2^-$  and OOH.

The rate of production of superoxide in a xanthine oxidase system was determined as outlined by Fridovich.<sup>6</sup> Briefly, the change in absorbance of cytochrome  $c(\Delta\epsilon_{550}=2.1\times10^4~M^{-1}~cm^{-1})$  was followed in a system containing 0.5 mM hypoxanthine, 0.1 mM cytochrome c, 50  $\mu$ M DETAPAC, and xanthine oxidase ( $\approx 0.25$ –20 mU/ml) in 50 mM phospate buffer, pH 7.4.

The spin trapping incubations used to determine [DMPO/OOH]<sub>ss</sub> contained 0.10 M DMPO, 0.5 mM hypoxanthine,  $50 \,\mu\text{M}$  DETAPAC, and varying amounts of X.O. such that the rate of  $O_2^-$  production varied from 9–71 nMs<sup>-1</sup>. These X.O. concentrations produced a constant rate of superoxide production in the time range of 3–8 minutes after the introduction of X.O. and an apparent steady-state concentration of DMPO/OOH as determined by repetative scans of the high field doublet of

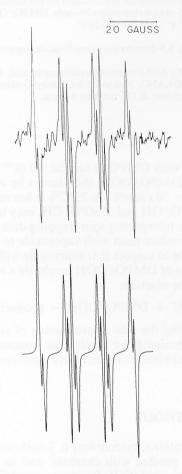


FIGURE 1 Top: The superoxide spin adduct spectrum of DMPO produced by a solution of 0.1 mM xanthine, 150  $\mu$ M DETAPAC, 70 mM DMPO and  $\approx 12$  mU/ml of X.O. in 50 mM phosphate buffer, pH 7.8. Instrument settings were: power, 20 mW; scan rate 25 G min<sup>-1</sup>; modulation amplitude, 0.3 G, time constant 0.5 s. *Bottom*: Simulation of DMPO/OOH spectrum assuming two species are present at equal population. The parameters used were:  $a_N^1 = 14.25$  G,  $a_H^1 = 12.45$  G and  $\Delta H_{pp}^1 = 0.96$  G,  $a_N^2 = 14.25$  G,  $a_H^2 = 10.10$  G. and  $\Delta H_{pp}^2 = 1.11$  G. A 50% Lorentzian-50% Gaussian shape function was used.

TABLE 1

rate <sub>1</sub> /nMs <sup>-1</sup>	[DMPO/ OOH] <sub>ss</sub> /nM	$k_5/10^6 \text{ M}^{-1} \text{ s}^{-1}$
9.1	212	7.2
13.8	285	5.8
21.2	357	5.2
40	590	3.4
71	765	2.9
	$k_5 = 4.9(\pm 2.2) \times 10^6 \mathrm{M}^{-1} \mathrm{s}^{-1}$	

The data of columns 1 and 2 represent the median of at least three determinations.

Use of the steady state assumption that  $d[O_2^-]/dt = 0$  and d[DMPO/OOH]/dt = 0 allows an exact solution for the unknown rate constant,  $k_5$ .

As seen in Table 1, the second-order rate constant determined with this kinetic model is  $\sim 5 \times 10^6 \, \text{M}^{-1} \, \text{s}^{-1}$ . This high rate constant implies that DMPO/OOH can compete effectively for  $O_2^-$  in the typical superoxide spin trapping experiment that uses DMPO(0.1 M). For example, the  $[O_2^-]_{ss}$  in the experiment where rate<sub>1</sub> = 21.2 nM s<sup>-1</sup> was calculated to be 4.4 nM. Thus, at pH 7.4 rate<sub>3</sub> = 13.2 nM s<sup>-1</sup> while rate<sub>5</sub> = 8.2 nM s<sup>-1</sup> and rate<sub>4</sub> = 4.9 nM s<sup>-1</sup>, i.e., the loss of DMPO/OOH due to self-decay is only one half that due to the superoxide-induced decay. That rate<sub>5</sub> is greater than rate<sub>4</sub> accounts for the low levels of DMPO/OOH seen in spin trapping experiments.

If in this kinetic model k5 were zero, then

$$k_3[DMPO][O_2^-]_{ss} = k_4[DMPO/OOH]_{ss}$$

After calculating  $[O_2^-]_{ss}$  (7.1 nM) we have  $[DMPO/OOH]_{ss} = 1.5 \,\mu\text{M}$ , but I only observed 0.36  $\mu$ M DMPO/OOH. Therefore, this value of  $k_5$  accounts for the less than predicted concentration of DMPO/OOH seen in spin trapping experiments.

These results point to the difficulty in attempting to do quantitative work by observing DMPO/OOH. In addition, it may be an experimental advantage in some experiments to arrange a low rate of superoxide generation so that reaction (5) can be minimized. Then, use of a slower scan rate with longer time constant or signal averaging can be employed because oxygen depletion will be delayed significantly.

A close examination of the results presented in Table 1 reveals that as the rate of superoxide generation increases,  $k_5^{obs}$  decreases. This trend suggests that the kinetic model may not be complete. The lower values of  $k_5^{obs}$  at higher  $O_2^-$  fluxes, i.e., high X.O. concentrations, suggest that a relative increase in DMPO/OOH concentration occurs. This would not be the case if X.O. were reducing DMPO/OOH. (Samuni *et al.*<sup>4</sup> found no evidence that X.O. directly destroys DMPO/OH.) A relative increase in [DMPO/OOHJ]<sub>ss</sub> would occur if the diamagnetic products of reaction (5) could be reoxidized to DMPO/OOH by superoxide, i.e.,

$$***[O_2^-]_{ss} \ = \ \frac{2k_3[DMPO] \, - \, ((2k_3[DMPO])^2 \, - \, 4(-2k_2)(rate_1 \, + \, k_4[DMPO/OOH]))^{1/2}}{2(-2k_2)}$$

and

$$k_5 \ = \ \frac{k_4 [DMPO/OOH] \ - \ k_3 [DMPO][O_2^-]}{-[DMPO/OOH][O_2^-]}$$

DMPO/OOH. (The high field doublet was chosen because it is least affected by the DMPO/OH signal that is also present.)

The determination of the concentration was accomplished by using 3-carboxy-proxyl as a standard. Because there are many factors that affect ESR signal area measurements, the 3-CP standard and DMPO/OOH experimental spectra were obtained using identical instrument settings, except for receiver gain, and identical physical arrangement of the samples in the cavity: Linearity of the receiver gain was verified. Double integration of the spectra was accomplished with the aid of the free radical simulation program of Oehler and Janzen. The relative area for each species was found by simulation of the midfield line of 3-CP and the high field doublet of DMPO/OOH. The 3-CP lineshape was simulated using  $\Delta$ Hpp = 1.275 G and a 70% Gaussian-30% Lorentzian shape function. The high field doublet of DMPO/OOH was simulated using  $a^{H} = 1.15$  G,  $\Delta$ Hpp = 1.11 G for the low field component and  $\Delta$ Hpp = 0.96 G for the high field component. For each component a 50% Gaussian-50% Lorentzian shape function was used.

The DMPO/OOH ESR spectrum has an asymmetry that traditional simulation efforts do not reproduce. However, the asymmetrical DMPO/OOH spectrum can be reproduced when two species of equal population, but different line widths, are used. An excellent fit is obtained if for species 1,  $a_N^1 = 14.25\,G$ ,  $a_H^1 = 12.45\,G$  and  $\Delta H_{pp}^1 = 0.96\,G$  for species 2,  $a_N^2 = 14.25\,G$ ,  $a_H^2 = 10.10\,G$  and  $\Delta H_{pp}^2 = 1.11\,G$ . See figure 1. These parameters showed that for DMPO/OOH and 3-CP lines of equal height, the relative area for the two species is: (area DMPO/OOH)/(area 3-CP) = 2.9. This information allows the calculation of the DMPO/OOH concentration from ESR signal height measurements using 3-CP as a standard. ESR spectra were recorded using a Varian E-4 system.

#### RESULTS AND DISCUSSION

To estimate the rate of the reaction of superoxide with DMPO the following system of kinetic equations was used:

$$O_2$$
 + hypoxanthine  $\xrightarrow{X.O.}$   $O_2^-$  + other products  
rate<sub>1</sub> measured with cytochrome c<sup>5</sup> (1)

$$O_2^- + O_2^- + 2H^+ \longrightarrow O_2 + H_2O_2$$
  
 $k_2^{\text{obs}} = 2.4 \times 10^5 \,\text{M}^{-1} \,\text{s}^{-1}; \,\text{pH} \, 7.4^9$  (2)

$$O_2^- + DMPO + H^+ \longrightarrow DMPO/OOH$$
  
 $k_3^{obs} = 30 M^{-1} s^{-1}; pH 7.4^1$ 
(3)

DMPO/OOH 
$$\longrightarrow$$
 products  
 $k_4^{\text{obs}} = 1.4 \times 10^{-2} \text{ s}^{-1}$ ; pH 7.4<sup>2\*\*</sup>

DMPO/OOH + 
$$O_2^- \longrightarrow \text{products}$$
  
 $k_5^{\text{obs}} = ?$  (5)

<sup>\*\*</sup>It has been reported the DMPO/ OOH decomposes with a half-life of 8 minutes. <sup>10</sup> However, this claim could not be substantiated using the riboflavin-DETAPAC system; <sup>2</sup> rather, at pH 7.4 I found a first-order half-life of 50 s, as previously reported. <sup>2</sup>

products<sub>5</sub> 
$$\frac{\text{slow}}{\text{k.c is small}}$$
 irreversible ESR-silent products (6)

$$products_5 + O_2^{-} + 2H^{+} \xrightarrow[k_7 \text{ is large}]{\text{fast}} DMPO/OOH + H_2O_2$$
 (7)

In essence, reaction (7) could occur if the initial ESR-silent products of reaction (5) could be efficiently reoxidized by  $O_2^-$ /OOH. This possibility has precedent. It has recently been demonstrated that the nitroxide/hydroxylamine couple of OXANO/OXANOH undergoes a reaction sequence parallel to reactions (5) and (7) above. However, Samuni *et al.* were not able to reoxidize the ESR-silent product of DMPO/OH +  $O_2^-$  with either ferricyanide or  $H_2O_2/Cu(II)$ . If reactions (6) and (7) were to be included in the reaction scheme, then gathering the experimental data for an exact solution becomes a problem. If reactions (6) and (7) are operative, then the value of  $k_5$  at pH 7.4 is probably  $\approx 10^7 \, M^{-1} \, s^{-1}$ . Nonetheless, even with the kinetic model used to arrive at  $k_5^{obs}$ , the value of  $5 \times 10^6 \, M^{-1} \, s^{-1}$  is a very useful number that can be used as a guideline for researchers to help interpret spin trapping data dealing with DMPO/OOH.

## References

- Finkelstein, E., Rosen, G.M. and Rauckman, E.J. Spin trapping. Kinetics of the reaction of superoxide and hydroxyl radicals with nitrones. J. Am. Chem. Soc., 102, 4994–4999, (1980).
- 2. Buettner, G.R. and Oberley, L.W. Considerations in the spin trapping of superoxide and hydroxyl radical in aqueous systems using 5,5-dimethyl-1-pyrroline-1-oxide. *Biochem. Biophys. Res. Commun.*, **83**, 69–74, (1978).
- 3. Samuni, A., Black, C.D.V., Krishna, C.M., Malech, H.L. Bernstein, E.F. and Russo, A. Hydroxyl radical production by stimulated neutrophils reappraised. *J. Biol. Chem.*, **263**, 13797–13801, (1988).
- Samuni, A., Krishna, C.M., Riesz, P., Finkelstein, E. and Russo, A. Superoxide reaction with nitroxide spin-adducts. Free Rad. Biol. Med., 6, 141–148, (1989).
- 5. Buettner, G.R. In the absence of catalytic metals ascorbate does not autoxidize at pH 7: ascorbate as a test for catalytic metals. *J. Biochem. Biophys. Meth.*, 16, 27-40, (1988).
- Fridovich, I. Cytochrome c. In Handbood of Methods for Oxygen Radical Research (ed. R.A. Greenwald), CRC Press, Boca Raton, pp 121–122, (1985).
- 7. Eaton, S.S. and Eaton, G.R. Signal area measurements in EPR. Bull. Mag. Reson., 1, 130-138, (1980).
- 8. Oehler, U.M. and Janzen, E.G., Simulation of isotropic electron spin resonance spectra: a transportable basic program. *Can. J. Chem.*, **60**, 1542–1548, (1982).
- Bielski, B.H.J., Cabelli, D.E. and Arudi, R.L., Reactivity of HO<sub>2</sub>/O<sub>2</sub><sup>-</sup> radicals in aqueous solution. J Phys Chem. Ref. Data., 14, 1041–1100, (1985).
- Turner, M.J., III and Rosen, G.M., Spin trapping of superoxide and hydroxyl radicals with substituted pyrroline 1-oxides. J. Med. Chem., 29, 2439-2444, (1986).
- Samuni, A., Krishna, C.M., Riesz, P., Finkelstein, E. and Russo, A. A novel metal-free low molecular weight superoxide dismutase mimic. J. Biol. Chem., 263, 17921–17924, (1988).

Accepted by Prof. E.G. Janzen