

This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2005)

offered by the

Free Radical and Radiation Biology Program

B-180 Med Labs

The University of Iowa

Iowa City, IA 52242-1181

Spring 2005 Term

Instructors:

GARRY R. BUETTNER, Ph.D.

LARRY W. OBERLEY, Ph.D.

with guest lectures from:

Drs. Freya Q . Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Paraquat, toxicity and mechanism

By

Changbin Du

Free Radical and Radiation Biology Program

Department of Radiation Oncology

The University of Iowa, Iowa city, IA, 52242-1181

For 77:222, Spring 2005

28 Feb 2005

Abbreviations:

GPx, glutathione peroxidase.

GSSG, glutathione disulfide.

H₂O₂, hydrogen peroxide.

HOCl, Hypochlorous acid

NADPH, nicotinamide adenine dinucleotide phosphate.

NO₂[•], nitrogen dioxide .

¹O₂, singlet oxygen

PQ²⁺, paraquat.

GSH, glutathione

GR, glutathione reductase

HO[•], hydroxyl radical.

MPO, myeloperoxidase.

NO[•], Nitric oxide

O₂^{-•}, superoxide.

ONOO⁻, Peroxynitrite.

PQ^{+•}, paraquat radical.

Outline	Page
Abstract.....	2
Introduction	3
The chemical and physical properties of paraquat	4
The toxicity of paraquat.....	4
Mechanism of paraquat toxicity.....	6
Summary.....	11
References.....	11

Abstract

Paraquat (1,1'-dimethyl-4,4'-bipyridinium dichloride) has been used for many years in many countries as a broad-spectrum herbicide. It is extremely toxic, causing multiple organ failure in humans. It preferentially damages the lungs, kidneys and liver, and may result in death. Paraquat is reduced by PQ^{2+} diaphorases or reducing agents. In aerobic condition, the reduced paraquat radical is reoxidized by oxygen, with formation of $O_2^{\cdot-}$, which in turn may be metabolized to other reactive oxygen species, including the highly reactive HO^{\cdot} , $ONOO^-$, or 1O_2 . Moreover, paraquat redox cycling leads not only to the generation of $O_2^{\cdot-}$, but also to a potential depletion of intracellular NADPH. The inability to maintain physiological levels of NADPH may cause cell damage. This review will focus on the physical and chemical properties of paraquat, its toxicity and the mechanism underlying it.

Introduction

Paraquat (PQ^{2+}) is one of the most widely used herbicides. It is used to control broad-leaved weeds and grasses, being less effective on deep-rooted plants such as dandelions. Paraquat does not harm mature bark, and is thus widely used for weed control in fruit orchards and plantation crops ^[A]. Paraquat is highly toxic to animals and has serious and irreversible delayed effects if ingested. The lungs selectively accumulate PQ^{2+} , and therefore contain higher concentrations than other tissues, which leads to fibrosis. Liver damage occurs and renal failure may follow as the kidneys remove absorbed PQ^{2+} [1]. Paraquat has also been used as a model factor inducing oxidative stress both *in vivo* and *in vitro* [2]. Although a definitive mechanism of toxicity of PQ^{2+} has not been delineated, it was proposed that a cyclic single electron reduction/oxidation of the PQ^{2+} molecule is a critical mechanistic event. Two consequences related to its toxicity are generation of reactive oxygen species and depletion of intracellular NADPH [3]. This review will focus on the physical and chemical properties of paraquat, its toxicity and the mechanism underlying it.

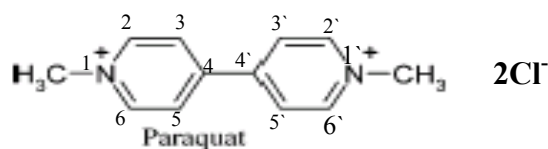


Figure. 1. Structure of herbicide paraquat dichloride.

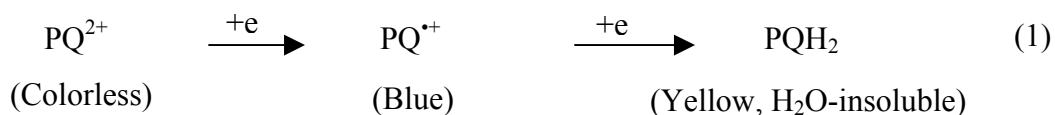
The basic bipyridyl consists of two pyridine rings linked together. N is counted as atom number-one in each ring [4].

^[A] <http://www.pan-uk.org/pestnews/actives/paraquat.htm> Accessed on 26/02/05.

The chemical and physical properties of paraquat

The herbicide paraquat is a quaternary nitrogen compound, whose basic chemical nucleus is bipyridyl consisting of two pyridine rings joined together. Their number-4 carbon atoms join the two pyridine rings. Each nitrogen atom has a methyl group (**Figure 1**) [4]. In its usually oxidized form, paraquat is ionized with two positive charges. So paraquat is usually manufactured as a salt with chloride ion (**Figure 1**).

Like other bipyridyl salts, paraquat is non-volatile both in the solid and in solvated state. Paraquat dichloride is extremely water-soluble and completely insoluble in non-polar organic solvents [5]. Paraquat is very stable in acid or neutral solutions. In alkaline solution paraquat decomposes to various complex colored degradation products [6]. The free radical forms of $PQ^{•+}$ are water-soluble and have a characteristic intensive color (**Reaction 1**). At 400 nm, the extinction coefficient for $PQ^{•+}$ is $4.6 \times 10^4 \text{ M}^{-1}\text{cm}^{-1}$, which allows the chemical detection of paraquat by spectrophotometry [9]. Paraquat has a redox potential of -446 mV [6], which allows the single electron reduction. **Table 1** summarizes some of the physical and chemical properties of paraquat.



Toxicology of paraquat

The extent of poisoning caused by paraquat depends on the amount, route, and duration of exposure and the person's condition of health at the time of the exposure. Paraquat

Table 1. Physical and chemical properties of paraquat [5].

Physical state	White (pure), yellow (technical), solid
Molecular weight	186.2 (Cation) 257.2 (dichloride)
Specific gravity	.24-1.26 20/20
Boiling point:	175 - 180°C ^[B]
Decomposition	300°C
Decompose	in ultraviolet light ^[C]
Vapor pressure	Nonvolatile
Density (g cm-3):	1.25 g/cm ³
Solubility at 20°C	Acetone—very slight Carbon disulfide—almost zero Dimethylformamide--slight Ethanol—extremely low Kerosene—almost zero Water—high soluble
Corrosiveness	Very to metals
Storage stability	Indefinitely long in original container
Incompatibilities	Alkyl sulfonate or alkyl aryl sulfonate Wetting agents, hydrolyzed by alkali Incompatible with strong oxidizing agents.

causes direct damage when it comes into contact with the lining of the mouth, stomach, or intestines^[D]. The lethal ingestion dose of paraquat in humans is 35 mg/kg^[E]. After paraquat enters the body, tissue distribution is ubiquitous with an apparent volume of distribution ranging from 1.2 to 1.6 l/kg [7]. In the research of Houze *et al*, the toxicokinetics of PQ²⁺ was studied in 18 cases of acute human poisoning using a specific radioimmunoassay. Plasma PQ²⁺ concentration exhibited a mean distribution half-life of 5 h and a mean elimination half-life of 84 h. Death related to pulmonary fibrosis occurred late and was associated with the elimination phase [7].

^[B] <http://ptcl.chem.ox.ac.uk/MSDS/PA/paraquat.html>. Accessed on 27/02/05.

^[C] <http://www.inchem.org/documents/jmpr/jmpmono/v070pr19.htm>. Accessed on 27/02/05.

^[D] <http://www.bt.cdc.gov/agent/paraquat/basics/facts.asp>. Accessed on 26/02/05.

^[E] <http://pmep.cce.cornell.edu/profiles/extoxnet/metiram-propoxur/paraquat-ext.html>. National Library of Medicine. Hazardous Substances Databank. Paraquat. 1992. Accessed on 09/03/05.

1. Immediate signs and symptoms of paraquat exposure

Ingestion of large amounts of paraquat can be extremely toxic. Symptoms such as pulmonary edema, lung scarring, liver failure, kidney failure, confusion, coma, seizures, injury to the heart, fast heart rate, muscle weakness, respiratory failure may happen within a few hours, possibly leading to death. Ingestion of small to medium amounts of paraquat may lead to adverse health effects like liver failure, kidney failure, heart failure and lung scarring after several days to weeks.

2. The long-term health effects

If a person survives the toxic effects of paraquat poisoning, long-term lung damage (scarring) is highly likely. Other long-term effects may also occur, including kidney failure, heart failure, and esophageal strictures^[D].

Mechanism of paraquat toxicity

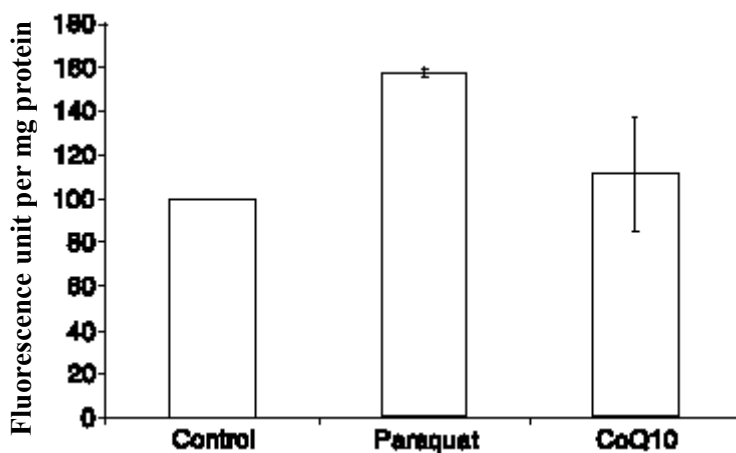
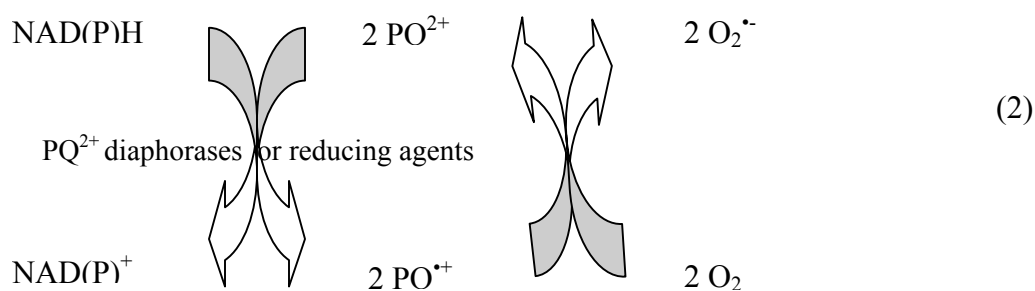


Figure. 2. Generation of reactive oxygen species. DCFDA was used to measure total cell ROS generation. After 48 h of paraquat treatment, human neuroblastoma SHSY-5Y cells showed a considerable increase in ROS production compared to control cells or cells pretreated with CoQ₁₀. Results were calculated per microgram

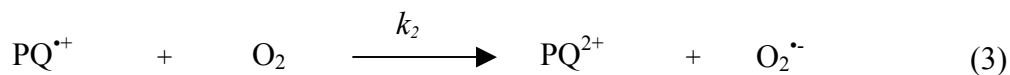
^[D] <http://www.bt.cdc.gov/agent/paraquat/basics/facts.asp>. Accessed on 26/02/05.

Produce Superoxide

Although a definitive mechanism of toxicity of paraquat has not been delineated, a cyclic single electron reduction/oxidation of the parent molecule is a critical mechanistic event [2]. Paraquat can induce oxidative stress and the following neuronal cell death (**Figure 2**) [7]. One-electron reduction of PQ^{2+} can be achieved by chemical reducing agents such as ascorbic acid (**Reaction 1**) [6, 13] or PQ^{2+} diaphorase [16]. PQ^{2+} diaphorase are usually oxidoreductase enzymes that contain flavins and use either NADH or NADPH as electron donors. A common cellular diaphorase that can redox cycle with PQ^{2+} is cytochrome P450 reductase (**Reaction 2**) [16]. So the oxidation of 1 mol of NADPH produces 2 mol of $O_2^{\bullet-}$.



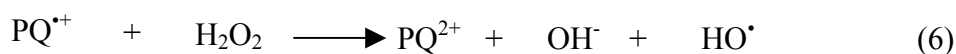
$PQ^{\bullet+}$ reacts very quickly with O_2 to give $O_2^{\bullet-}$ (**Reaction 3**), with $k_2 = 7.7 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$ [8].



After transfection of cDNA for human CuZn superoxide dismutase to NIH/3T3 cells, it becomes resistant to paraquat, which strongly supports the fact that formation of $O_2^{\bullet-}$ is a necessary part of its cytotoxic effects [10].

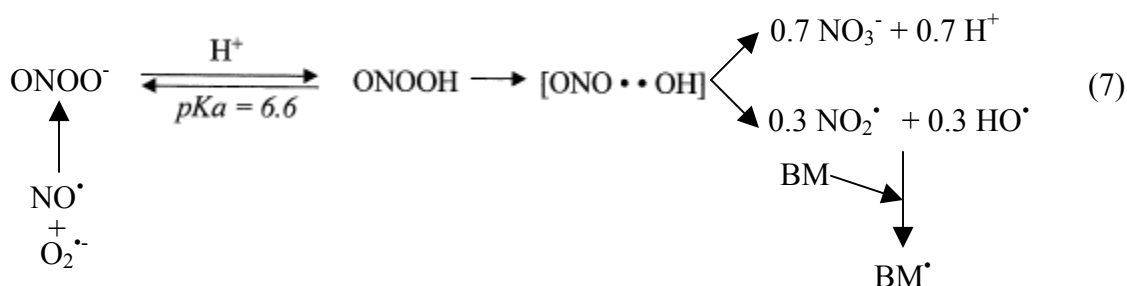
Produce Hydroxyl radical

Superoxide may be dismutated to H_2O_2 . In the presence of Fe^{2+} , highly reactive and toxic radicals such as HO^\bullet can be formed (**Reaction 4 & 5**).



In the research done by Winternourn, PQ^{+} can react with H_2O_2 to produce HO^\bullet directly in the absence of metal catalyst (**Reaction 6**) [13]. The reaction is fast and able to compete with the reaction of with O_2 . So production of HO^\bullet from PQ^{+} and H_2O_2 may therefore be of major significance in paraquat toxicity.

Produce ONOO^- and then HO^\bullet

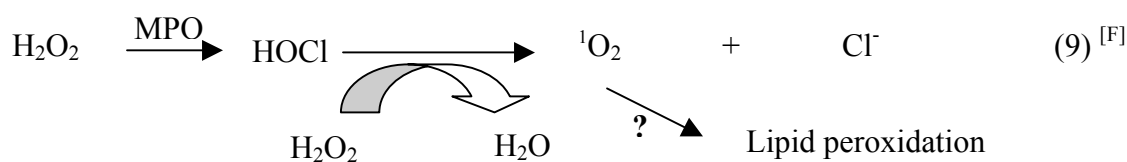


Nitric oxide (NO^\bullet) has been implicated in PQ^{2+} induced lung injury. It is thought to be due to its rapid reaction with PQ^{+} -formed $\text{O}_2^{\bullet-}$ to produce the strong oxidant peroxynitrite (**Reaction 7**) [14]. Peroxynitrite is a very reactive species, the “bent-form” of which has an E° of +2100 mV, which is nearly as oxidizing as HO^\bullet (+ 2310 mV). Peroxynitrite is

stable at alkaline pH but upon protonation ($pK_a = 6.6$) it decomposes rapidly ($k = 0.17 \text{ s}^{-1}$ at pH 7.4 and $k = 1.1 \text{ s}^{-1}$ at pH 5.4, 25°C) to yield 70% nitrate and 30% hydroxyl radical and nitrogen dioxide (NO_2^\bullet). Hydroxyl radical is capable of reacting with all biological macromolecules (lipids, proteins, nucleic acids and carbohydrates) to the corresponding radicals (**Reaction 7**) [15].

Produce Singlet Oxygen

Hara *et al* studied the PQ^{2+} -stimulated NADPH-dependent lipid peroxidation in mouse brain and pulmonary microsomes. They found that the lipid peroxidation was inhibited by SOD and singlet oxygen quenchers, but not by catalase or hydroxyl radical scavengers. These findings suggest that activated oxygen species, especially superoxide and singlet oxygen, may play a major role in the stimulation of microsomal lipid peroxidation by paraquat in both brain and lung [11]. In the absence of metal catalyst, hypochlorous acid (HOCl) can be generated enzymatically by myeloperoxidase (MPO) from H_2O_2 . Reaction of HOCl with H_2O_2 yields $^1\text{O}_2$ (**Reaction 9**)^[F].



The result of Hara *et al* is weird, however, it has been reported that singlet oxygen ($^1\text{O}_2$) can lead to lipid peroxidation [17]. Peroxidation of Linolenate has been reported by a photochemical source of singlet oxygen [18], however the mechanism is unclear.

^[F] http://www.rndsystems.com/asp/g_sitebuilder.asp?bodyId=222. Accessed on 10/03/05.

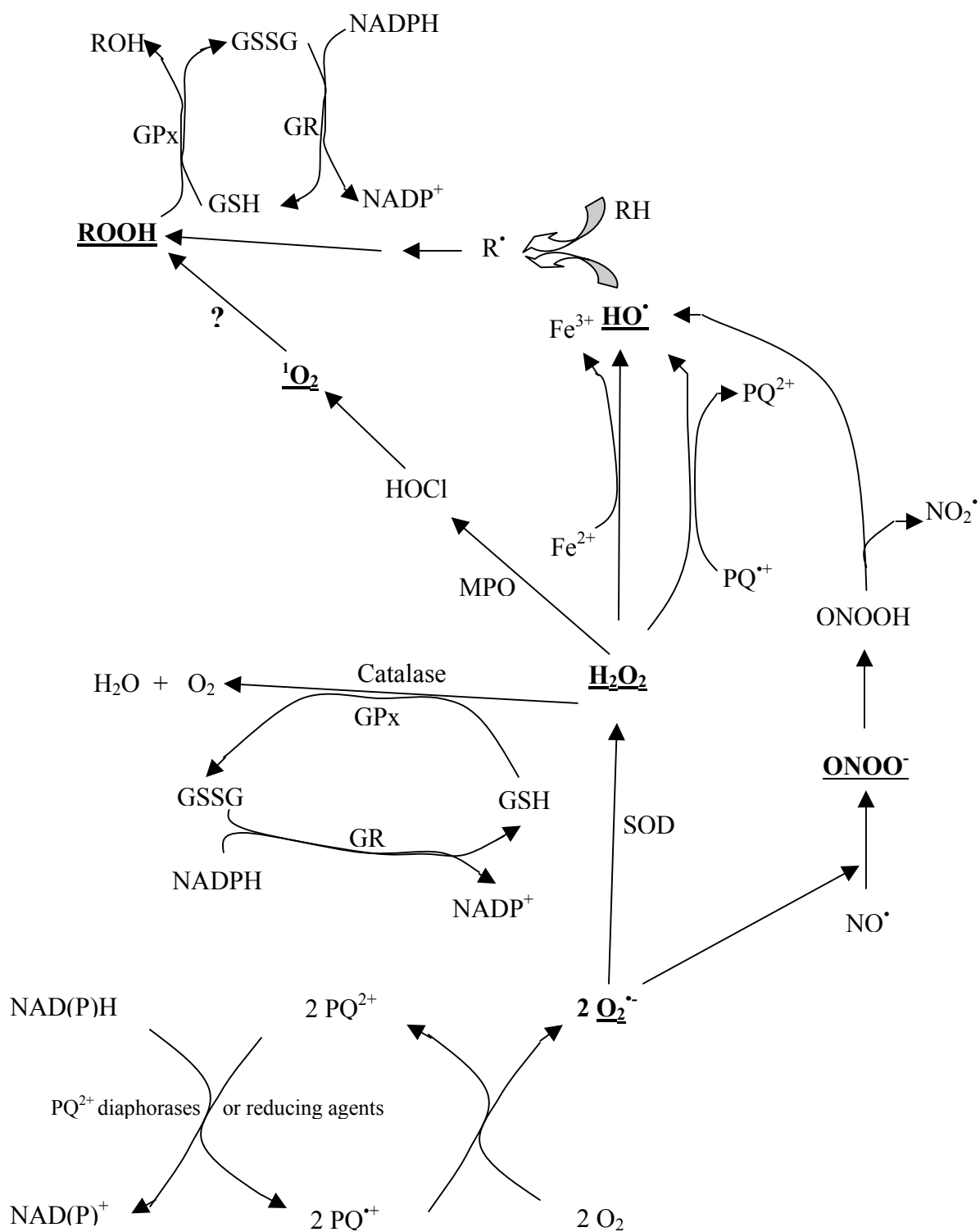


Figure 3. Proposed mechanism of action for the in vivo toxicity of paraquat. Important intermediate species or products are shown in bold and underlined.

Depletion of NADPH

Several enzymes are present in cells that can catabolize the $O_2^{\bullet-}$ and reduce lipid peroxidation. Superoxide is converted to H_2O_2 and O_2 by SOD, and H_2O_2 is further inactivated to H_2O and O_2 by catalase or glutathione peroxidase (GPx). The reduction of lipid peroxides by GPx requires glutathione (GSH). Because the reduction of glutathione disulfide (GSSG) is coupled with the oxidation of NADPH through glutathione reductase (GR), the availability of sufficient NADPH is a critical factor for the detoxification of paraquat. So the redox cycling of paraquat and the detoxification of $O_2^{\bullet-}$, H_2O_2 and lipid peroxidation consume NADPH. The inability to maintain physiological levels of NADPH may cause cell damage (**Figure 3**). Cells become more susceptible to free radical attack and peroxidation of vital cellular constituents.

Summary

Current reports suggest that the redox cycling, single-electron reduction/oxidation of the parent molecule is the critical event underlying the toxicity of paraquat. Following production of reactive oxygen species and lipid production lead to loss of NADPH can cause cell death.

Reference

1. Czarniewska E, Kasprzyk A, Ziemnicki K. 2003. Effect of paraquat and metoxychlor on antioxidant enzymes in frog *Rana esculenta* L. liver. *Biol Lett* **40**: 25-133.
2. Bus JS, Gibson JE. (1984) Paraquat: Model for oxidant-initiated toxicity. *Environ Health Persp.* **55**:31-46

3. Rose MS, Smith LL, Wyatt I. (1976) The relevance of pentose phosphate pathway stimulation in rat lung to the mechanism of paraquat toxicity. *Biochem Pharmacol.* **25**:763-1767.
4. Bacigalupo MA, Meroni G, Mirasoli M, Parisi D, Longhi R. (2005) Ultrasensitive Quantitative Determination of Paraquat: Application to River, Ground, and Drinking Water Analysis in an Agricultural Area. *J Agric Food Chem.* **53**:216 – 219.
5. Haley TJ. (1979) Review of the toxicology of paraquat. *Clinical Toxicology* **14**:1-46
6. Calderbank A. (1968) The bipyridylum herbicides. *Advances Pest Contr Res.* **8**:127-235.
7. McCarthy S, Somayajulu M, Sikorska M, Borowy-Borowski H, Pandey S. (2004) Paraquat induces oxidative stress and neuronal cell death; neuroprotection by water-soluble Coenzyme Q10. *Toxicol Appl Pharmacol.* **201**:21-31
8. Houze P, Baud FJ, Mouy R, Bismuth C, Bourdon R, Scherrmann JM. (1990) Toxicokinetics of paraquat in humans. *Hum Exp Toxicol.* **9**:5-12.
9. Ledwith A. (1977) Electron transfers reactions of paraquat. In: Autor AP. ed. *Biochemical Mechanism of Paraquat Toxicity*. New York, San Francisco, London: Academic Press. pp: 21-38
10. Krall J, Bagley AC, Mullenbach GT, Hallewell RA, Lynch RE. (1988) Superoxide mediates the toxicity of paraquat for cultured mammalian cells. *J Biol Chem.* **263**:1910-1914.
11. Hara S, Endo T, Kuriwa F, Kano S. (1991) Mechanism of paraquat-stimulated lipid peroxidation in mouse brain and pulmonary microsomes. *J Pharm Pharmacol.* **43**: 731-733.
12. Shivhare P, Gupta VK. (1991) Spectrophotometric method for the determination of paraquat in water, grain and plant materials. *Analyst.* **116**: 391-393.
13. Winterbourn CC. (1981) Production of hydroxyl radicals from paraquat radicals and H₂O₂. *FEBS Lett.* **128**: 339-342.
14. Nemery, B. & van Klaveren, R. J. (1995). NO wonder paraquat is toxic. *Hum. Exp. Toxicol.* **14**: 308-309.
15. Augusto O, Bonini MG, Amanso AM, Linares E, Santos CC, X. and De Menezes SL. (2002) Nitrogen dioxide and carbonate radical anion: two emerging radicals in biology. *Free Radic Biol Med.* **32**:841-859.

16. Yamazaki I, Piette LH, Grover TA. (1990) Kinetic studies on spin trapping of superoxide and hydroxyl radicals generated in NADPH-cytochrome P-450 reductase-paraquat systems. *J Biol Chem.* **265**: 652-659.
17. Terao J. & Matsushita S. (1977) Products formed by photosensitized oxidation of unsaturated fatty acid esters. *J Am Oil Chem Soc* **54**: 234-239.
18. Kellogg EW 3rd, Fridovich I. (1975) Superoxide, hydrogen peroxide, and singlet oxygen in lipid peroxidation by a xanthine oxidase system. *J Biol Chem.* **250**: 8812-8817.