

This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2005)

offered by the

Free Radical and Radiation Biology Program

B-180 Med Labs

The University of Iowa

Iowa City, IA 52242-1181

Spring 2005 Term

Instructors:

GARRY R. BUETTNER, Ph.D.

LARRY W. OBERLEY, Ph.D.

with guest lectures from:

Drs. Freya Q. Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Tocopheroxyl Radical

by

KJERSTIN M. OWENS

Department of Free Radical and Radiation Biology
The University of Iowa
Iowa City, IA 52242-1181

For 77:222, Spring 2005

9. February 2005

Abbreviations

Asc^{•-} - Ascorbate Radical

AscH⁻ - Ascorbate

CoQ₁₀ - Ubiquinone-10

CoQ₁₀H^{•-} - Ubisemiquinone-10

CoQ₁₀H₂ - Ubiquinol-10

EGCG- (-)-Epigallocatechin gallate

EPR - Electron Paramagnetic Resonance

GSH - Glutathione

HO₂[•] - Hydroperoxyl Radical

LDL - Low Density Lipoprotein

LH - Lipid

LOO[•] - Lipid Peroxyl Radical

LOOH - Lipid Hydroperoxide

NRP - Nonradical product

O₂^{•-} - Superoxide

TMP - Tocopherol-Mediated peroxidation

TO[•] - Tocopheroxyl Radical

TOH - Tocopherol

Outline

I.	Introduction	3
II.	Oxidation of tocopherol to form tocopheroxyl radical	4
	A. “Conventional” LDL lipid peroxidation	5
	B. Tocopherol-mediated peroxidation	5
III.	Reduction of tocopheroxyl radical	6
	A. Ascorbate	7
	B. Ubiquinone-10	7
IV.	Vitamin E and disease	8
V.	Conclusion	8
VI.	References	9

Abstract

Tocopheroxyl radical is a highly lipophilic compound that is produced as a result of the oxidation of tocopherol. Tocopherol is a potent chain-breaking antioxidant that is important in stopping lipid peroxidation. Two lipid peroxidation schemes involving both tocopherol and its radical propose similar yet very different roles for the antioxidant pair. In “conventional” LDL lipid peroxidation tocopherol acts as an antioxidant, whereas, in tocopherol-mediated peroxidation, tocopherol can be both a pro-oxidant and an antioxidant. The tocopheroxyl radical is able to react with other antioxidants and be recycled back into tocopherol, thus propagating the antioxidant properties of this compound. Two prominent recycling compounds of the tocopheroxyl radical are ascorbate and ubiquinol-10. The reduction of tocopheroxyl radical is important in maintaining antioxidant defenses.

Introduction

Vitamin E was discovered in 1922 by H.M. Evans at Berkeley University [1]. It was not until six decades later, in 1980 that Burton *et al.* recognized its role as a chain-breaking antioxidant [2]. Today, vitamin E is a popular dietary antioxidant. There are eight isoforms of vitamin E; four tocopherol and four tocotrienol structures [3]. The tocopherols (TOH) consist of a phenolic head and a long saturated carbon tail (**Figure 1**). They differ from tocotrienols in that tocotrienol's carbon chain is unsaturated, with three double bonds [3]. The tocopherol isomers give rise to the tocopheroxyl radical (TO[•]) by the oxidation of the hydroxyl group. This radical can be detected by both electron paramagnetic resonance (EPR) [4] and UV spectrometry [5] (**Figure 2**).

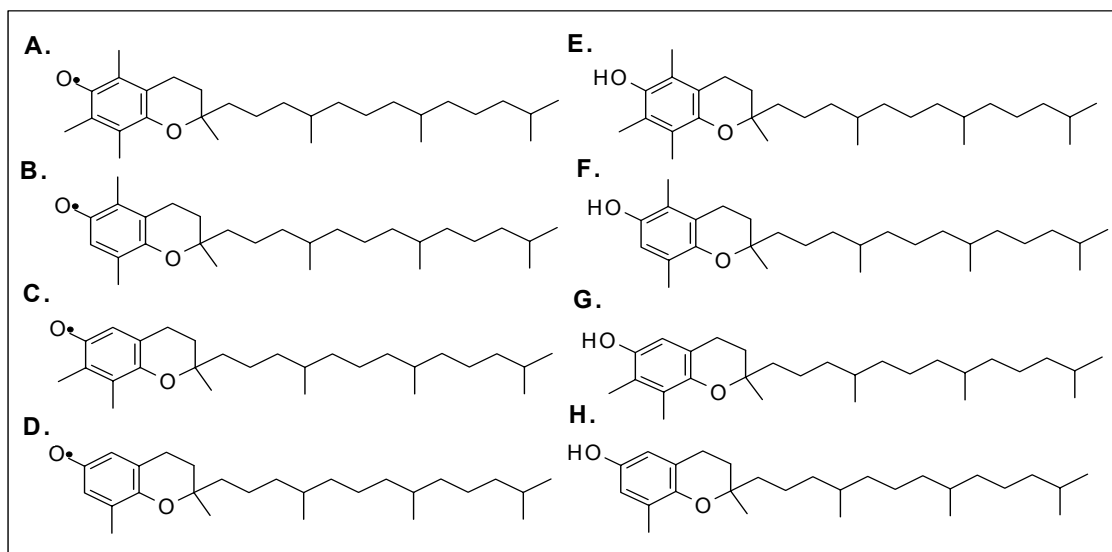


Figure 1. Structure of tocopheroxyl radicals and tocopherols. A. α -TO[•]. B. β -TO[•]. C. γ -TO[•]. D. δ -TO[•]. E. α -TOH. F. β -TOH. G. γ -TOH. H. δ -TOH

Because of their long carbon chain tocopherols and tocotrienols and their radicals are very hydrophobic [6]. This allows them to reside in lipid rich areas, particularly membranes and in LDL, where they act to stop lipid peroxidation. The lipophilic nature of this radical creates

some problems with its accessibility to other antioxidants. This paper will discuss the formation of TO^{\bullet} through the oxidation of TOH and the reduction of the radical back into its antioxidant form.

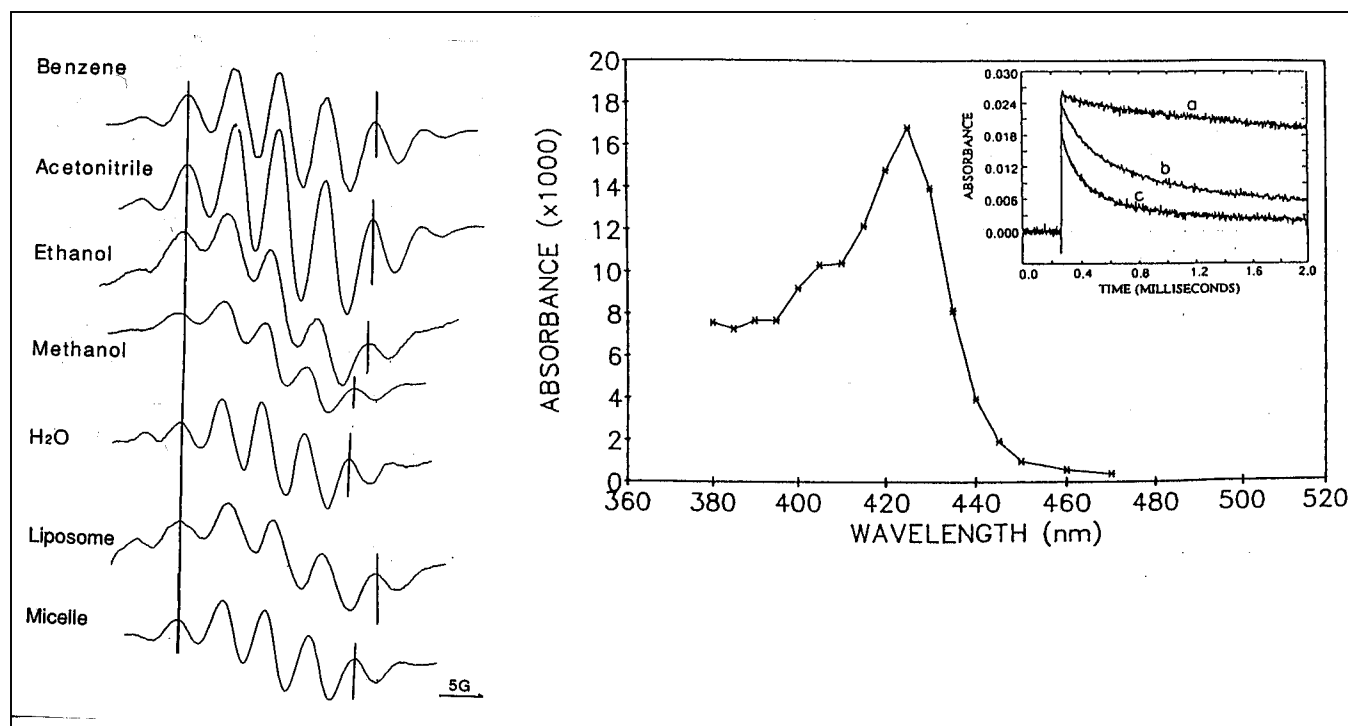


Figure 2. Detection of TO^{\bullet} . A. EPR spectra of TO^{\bullet} in several different solutions [4]. UV spectrum of TO^{\bullet} . Inset. Decay of TO^{\bullet} at 425 nm at varying pH (a. pH 6.6; b. pH 4.2; c. pH 3.2) [5].

Oxidation of Tocopherol to Form Tocopheroxyl Radical

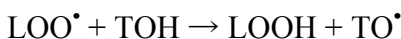
Phenols are typically chain-breaking antioxidants [7]. Tocopherol is a chain breaking antioxidant that is able to stop lipid peroxidation. In doing so, its hydroxyl group of the TOH head is oxidized to become TO^{\bullet} [1]. If not stopped, lipid peroxidation in membranes can reach 1 to 5 nmol mg^{-1} protein min^{-1} [1]. TOH is present in the membrane at a ratio of 1 to 2000 phospholipids [1,8]. Even at this small concentration (less than 0.05 to 0.1 nmol mg^{-1} protein [1]), TOH is able to protect the membrane against lipid peroxidation.

All forms of vitamin E are able to stop lipid peroxidation in this manner. However, there is a hierarchy to the efficiency of the isomers. α -TOH is the best chain-breaking antioxidant of

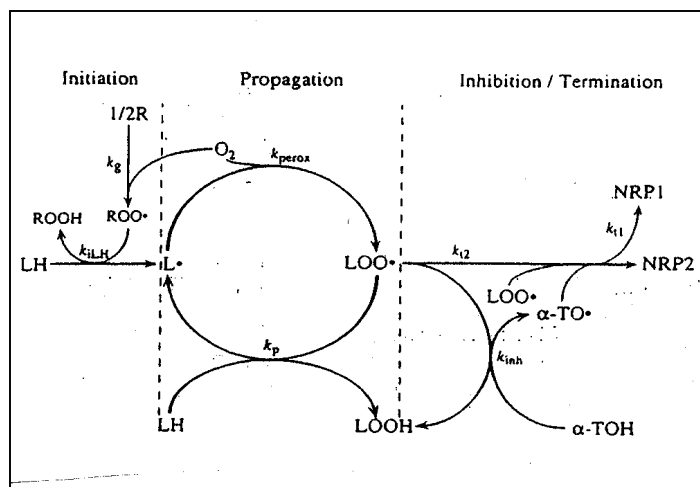
the isomers with a rate constant of $2.4 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$, followed by β -TOH ($1.7 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$), γ -TOH ($1.6 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$), and finally δ -TOH ($6.5 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$) [6]. α -TOH also is the most abundant isomer of vitamin E found in mammalian membranes [8,9].

“Conventional” LDL Lipid Peroxidation

In “conventional” lipid peroxidation, TOH acts only as an antioxidant to stop lipid peroxidation (**Figure 3**). The radical reaction is started with an initiation event in which a hydroperoxyl causes the oxidation of a polyunsaturated fatty acid. This reacts with oxygen to forms LOO^\bullet . This is carried out in the propagation step where an uncertain number of peroxidation cycles take place. In the inhibition (or termination) step, TOH donates a hydrogen atom to LOO^\bullet and is oxidized to form TO^\bullet , thus stopping the chain of peroxidation [1,9,10,11].



When TO^\bullet is recycled back in to TOH, TOH can act to stop another propagation of lipid peroxidation. However, TO^\bullet may react with LOO^\bullet to form a nonradical product (NRP) [9,10].

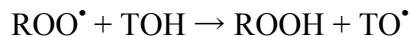


Tocopherol-Mediated Peroxidation

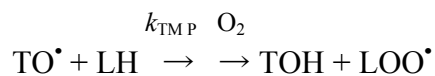
In tocopherol-mediated peroxidation, as proposed by Stocker *et al.* [9-12], TOH can act as either a pro-oxidant or an antioxidant (**Figure 4**). In this case TOH transfers a peroxidation

Figure 3. “Conventional” LDL lipid peroxidation. The radical then oxidizes LH to begin the cycle of lipid peroxidation. This step cycles as a part of propagation to create an indefinite amount of LOO^\bullet . Inhibition or termination occurs when TOH reduces LOO^\bullet to LOOH , forming TO^\bullet . Alternatively, LOO^\bullet may react with itself or TO^\bullet to form a nonradical product [9].

reaction from the aqueous phase into the lipid phase, shown in the following equation [11].



The radical reaction is now facilitated within the lipid environment *via* the oxidation of LH by TO^\bullet [12].



The rate constant for this reaction (k_{TMP}) is approximately $0.1 \text{ M}^{-1} \text{ s}^{-1}$ [12]. Although this reaction is very slow, TO^\bullet must wait for several minutes before being recycled back to TOH by another antioxidant. This provides ample time for TMP to occur [11,12]. Lipid peroxidation is thus initiated by TO^\bullet and is then terminated by TOH. This creates a cycle of tocopherol-mediated peroxidation [7,9-11].

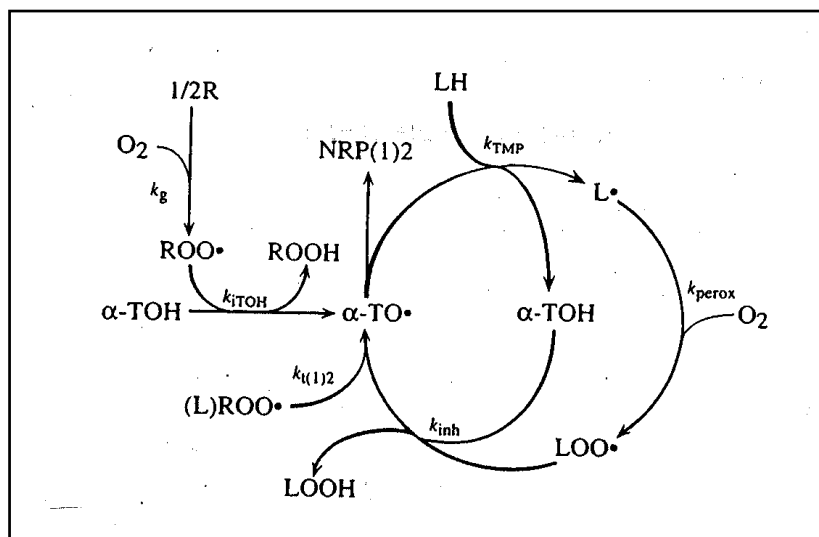


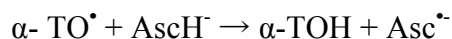
Figure 4. Tocopherol-mediated peroxidation cycle. Hydroperoxyl radicals oxidize TOH to TO^\bullet . Tocopheroxyl radical then acts as a pro-oxidant, oxidizing LH, which forms L^\bullet . The addition of oxygen to makes LOO^\bullet which reacts with TOH to reform TO^\bullet and restart the cycle of peroxidation [9].

Reduction of Tocopheroxyl Radical

The tocopheroxyl radical can be recycled back into tocopherol by various antioxidants. Many antioxidants are able to reduce TO^\bullet , such as ascorbate [10,12-15,19], ubiquinol-10 [11,16,17], GSH [1,8,12], EGCG [18], carotenoids [19], BHT [10], and even TOH [2]. Ascorbate and ubiquinol-10 are described in more detail below.

Ascorbate

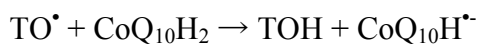
Ascorbate (AscH⁻) goes through a one-electron reduction of TO[•] to regenerate TOH [5,19].



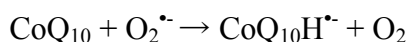
In solution (water/isopropanol/acetone), the rate constant for the reduction of TO[•] by ascorbate is $1.55 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ [14]. In a phosphatidylcholine liposome the rate constant increases to $2 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ [12,14,15]. Because AscH⁻ is hydrophilic, it is not able to be in the membrane component as is the highly hydrophobic TO[•]. The radical center of TO[•] is at the lipid-aqueous interphase of the membrane [9]. Here, the AscH⁻ is able to react with the unpaired electron of TO[•]. This reduction moves the radical center from the hydrophobic membrane to the aqueous phase so the radical can no longer react with LH [12]. Because Asc^{•-} is resonance-stabilized, it is relatively unreactive and is unlikely to propagate more free radical reactions [7].

Ubiquinol-10

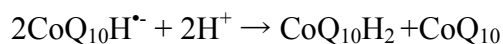
Ubiquinone (Coenzyme Q, CoQ₁₀) is an electron carrier in the electron transport chain. Because of its position in the mitochondrial membrane, ubiquinol-10 (fully reduced ubiquinone) is able to react with TO[•] and reduce it back to TOH [16].



The second order rate constant for this reaction was found to be $3.74 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ in benzene and $2.15 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ in ethanol [16]. While TOH can be oxidized by protonated superoxide ($k = 2 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$), superoxide also acts as an ally by helping to reduce TO[•] [16]. It does so by a one-electron reduction reaction of CoQ₁₀ to form ubisemiquinone-10 radical [17].



Two ubisemiquinone-10 radicals react to form ubiquinol-10, which then can reduce TO^\bullet as shown above [17].



Vitamin E and Disease

When vitamin E was first discovered it was known to be essential for reproduction [2, 20]. Since then its biological implications have gained great importance. In arterial walls TOH stops the oxidation of LDL, which heads off atherosclerosis by preventing the recruitment of macrophages to ingest the LDL and for the foam cells of an atherosclerotic lesion [9,10]. TOH also prevents oxidation of polyunsaturated fatty acids in synaptosomes, which may play a key preventative role in Parkinson's disease [21]. Finally, TOH antioxidant properties help protect against the oxidative stresses that are a factor in carcinogenesis [21]. As discussed earlier, the properties of tocopheroxyl radical are an important part of vitamin E cycling that allows tocopherol to continuously do its job.

Conclusion

Tocopheroxyl radical is an important factor in free radical chemistry on lipid peroxidation. According to "conventional" LDL peroxidation TOH acts only as an antioxidant and the only role for TO^\bullet is to be reduced back into TOH [1,9-11]. However, tocopherol-mediated peroxidation, a more recent model, suggests that TOH is a pro-oxidant and an antioxidant in lipid peroxidation, and TO^\bullet help propagate the radical reactions [9-12]. This model increases the emphasis on the importance of ascorbate and ubiquinol-10 in the antioxidant system.

References

1. Packer L. (1992) New horizons in vitamin E research-the vitamin E cycle, biochemistry, and clinical applications. In: Ong ASH, Packer L, ed. *Lipid-Soluble Antioxidants: Biochemistry and Clinical Applications*. Basel, Switzerland: Birkhäuser Verlag; pp. 1-16.
2. van Acker SABE, Koymans LMH, Bast A. (1993) Molecular pharmacology of vitamin E: Structural aspects of antioxidant activity. *Free Radic Biol Med.* **15**: 311-328.
3. Servinova E, Kagan V, Han D, Packer L. (1991) Free radical recycling and intramembrane mobility in the antioxidant properties of alpha-tocopherol and alpha-tocotrienol. *Free Radic Biol Med.* **10**: 1-15.
4. Iwatsuki M, Tsuchiya J, Komuro E, Yamamoto Y, Niki E, (1994) Effects of solvents and media on the antioxidant activity of α -tocopherol. *Biochim Biophys Acta.* **1200**: 19-26.
5. Bisby RH, Parker AW. (1995) Reaction of ascorbate with the α -tocopheroxyl radical in micellar and bilayer membrane systems. *Arch Biochem Biophys.* **317**: 170-178.
6. Bisby RH, Parker AW. (1993) Radiation-induced free radical reactions. In: Ong ASH, Packer L, ed. *Free Radicals: From Basic Science to Medicine*. Basel, Switzerland: Birkhäuser Verlag; pp. 31-37.
7. Burton GW, Cheeseman KH, Doba T, Ingold KU, Slater TF. (1983) Vitamin E as an antioxidant *in vitro* and *in vivo*. In: Porter R, Whelan J, ed. *Biology of Vitamin E*. London, Eng: Pittman; pp. 4-18.
9. Waldeck AR, Stocker R. (1996) Radical-initiated lipid peroxidation in low density lipoprotein: Insights obtained from kinetic modeling. *Chem Res Toxicol.* **9**: 954-964.
8. Maguire JJ, Wilson DS, Packer L. (1989) Mitochondrial electron transport linked tocopheroxyl radical reduction. *J Biol Chem.* **264**: 21462-21465.
10. Bowry VW, Stocker R. (1993) Tocopherol-mediated peroxidation. The prooxidant effect of vitamin E on radical-initiated oxidation of human low-density lipoprotein. *J Am Chem Soc.* **115**: 6029-6044.
11. Thomas SR, Neuzil J, Stocker R. (1997) Inhibition of LDL oxidation by ubiquinol-10. A prospective mechanism for coenzyme Q in atherogenesis? *Mol Asp Med.* **18**: s85-s103.
12. Bowry VW, Mohr D, Cleary J, Stocker R. (1995) Prevention of tocopherol-mediated peroxidation in ubiquinol-10 free human low density lipoprotein. *J Biol Chem.* **270**: 5756-5763.
13. McCay PB. (1985) Vitamin E: Interactions with free radicals and ascorbate. *Ann Rev Nutr.* **5**: 323-340.

14. Mukai K, Nishimura M, Ishizu K, Kitaman Y. (1989) Kinetic study of the reaction of vitamin C with vitamin E radicals (tocopheroxyls) in solution. *Biochim Biophys Acta*. **991**: 276-279.
15. Mehlhorn RJ, Sumida S, Packer L. (1989) Tocopheroxyl radical persistence and tocopherol consumption in liposomes and in vitamin E-enriched rat liver mitochondria and microsomes. *J Biol Chem*. **264**: 13448-13452.
16. Mukai K, Kikuchi S, Urano S. (1990) Stopped-flow kinetic study of the regeneration reaction of tocopheroxyl radical by reduced ubiquinone-10 in solution. *Biochim Biophys Acta*. **1035**: 77-82.
17. Stoyanovsky DA, Osipov AN, Quinn PJ, Kagan VE. (1995) Ubiquinone-dependent recycling of vitamin E radicals by superoxide. *Arch Biochem Biophys*. **323**: 343-351.
18. Zhou B, Wu LM, Yang L, Liu ZL. (2005) Evidence for α -tocopherol regeneration reaction of green tea polyphenols in SDS micelles. *FRBM*. **38**: 78-84.
19. Mortensen A, Skibsted LH. (1997) Relative stability of carotenoid radical cations and homologue tocopheroxyl radicals. A real time kinetic study of antioxidant hierarchy. *FEBS Lett*. **417**: 261-266.
20. Prasad KN, FACN, Edwards-Prasad J. (1992) Vitamin E and cancer prevention: recent advances and future potentials. *J Am Col Nutr*. **11**: 487-500.
21. Vatassery GT, Smith WE, Quach HT. (1994) Increased susceptibility to oxidation of vitamin E in mitochondrial fractions compared with synaptosomal fractions from rat brains. *Neurochem Int*. **24**: 29-35.