This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2001)

offered by the

Free Radical and Radiation Biology Program B-180 Med Labs The University of Iowa Iowa City, IA 52242-1181 Spring 2001 Term

Instructors: GARRY R. BUETTNER, Ph.D. LARRY W. OBERLEY, Ph.D.

with guest lectures from: Drs. Freya Q . Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Desferrioxamine

by

Michael L. Ott

311C Chemistry/Botany Building Department of Chemistry University of Iowa Iowa City, IA 52242

For 77:222, Spring 2001

12. February 2001

Abbreviations:

DFO- Desferrioxamine EPR- Electron paramagnetic resonance FAB-MS- Fast atom bombardment mass spectrometry HPLC- High pressure liquid chromatography M_w- Molecular weight UV/Vis- Ultraviolet/Visible spectrometry

Table of Contents

Abstract

Desferrioxamine is a chelator of iron, aluminum and other metals. It is a hexadentate ligand that binds with an extremely favorable stability constant. This property of desferrioxamine makes it ideal for treating diseases such as thassalemia, in which the body is overloaded with iron. Desferrioxamine can also be oxidized to give a nitroxide radical, which leads to some interesting chemistry. The properties, reactions and biological activities of desferrioxamine will be discussed.

Introduction

Iron is essential for life, but as is often the case, it is possible to get too much of a good thing. If that happens, desferrioxamine could be used to sponge up the excess. Desferrioxamine is a hexadentate chelating ligand, which binds to Fe^{3+} with a stability constant near 10^{31} [1]. It is somewhat selective, in that it binds to Fe^{2+} with a stability constant of 10^7 [2] and other first row transition metals in the range of 10^{10} to 10^{15} [3]. Desferrioxamine is a large molecule, composed of one acetic acid, two succinic acids, and three molecules of 1-amino-5-hydroxylaminopentane, giving a M_w of 560 a.u. Desferrioxamine was first extracted from *Streptomyces pilosus*, where it was a natural iron chelator. Its basic character and ability to form inorganic salts derives from its terminal free amino group [4], and desferrioxamine has a pK_a of 9.2 [5].

When desferrioxamine is combined with iron, it is called ferrioxamine. To further complicate matters, desferrioxamine is referred to by many names in the literature. Some are alternate spellings: desferioxamine, deferrioxamine, and deferioxamine, while others are totally different, like DesferaTM. Because the molecule is not ferocious or savage, it is not called feral when combined with iron.

The structure of desferrioxamine is shown below, in both chain form and chelated to Fe³⁺.



Figure 1- Above, desferrioxamine in the chain form, and, at right, ferrioxamine chelated to Fe^{3+} . Notice the complicated arrangement around the Fe atom. Adapted from [4].



Stability of desferrioxamine complexes

Desferrioxamine is selective towards Fe^{3+} because of their extremely favorable binding constant, nearly 10^{31} . The binding constants of other metals are shown below, in Table 1. This was adapted from [18].

Metal	Fe ³⁺	Fe ²⁺	Ni ²⁺	Cu ²⁺	Zn^{2+}	Cd^{2+}	Al^{3+}	La ³⁺	Yb ³⁺
Stability Constant	30.7	7.2	10.9	14.1	10.1	7.9	23.1	10.9	16.0
[M-DFO]/[M][DFO]									
(log K =)									

The most stable compound, by far, is Fe^{3+} -DFO. Its nearest neighbor is Al^{3+} -DFO. The preference of DFO for Fe^{3+} rather than Fe^{2+} is clearly shown above.

Desferrioxamine in biological systems

There are several biological uses for desferrioxamine. Thassalemia is a disease caused by iron overload, and a series of subcutaneous desferrioxamine injections are the primary treatment [6]. Free radicals are produced during the ischemia/reperfusion sequence of all open-heart operations. Transition metals are responsible for some of the radicals produced, and DFO can inhibit their action [7]. A DFO derivative, hydroxyethyl-starch desferrioxamine, has been used to delay the onset of diabetes mellitus in rats. Once again, the mechanism was believed to involve the chelation of iron, but only seemed to work on male rats. Further studies are ongoing to elucidate this discrepancy [8].

Desferrioxamine can also be used to help cells survive high levels of H_2O_2 . When DFO is present, enough iron is chelated to prevent the harmful breakdown of H_2O_2 , and

cells could deal with it by normal means, such as catalase [9]. Iron isn't the only metal chelated by DFO. The presence of DFO-Mn(III), an SOD mimic, inhibits cataract formation in rabbits by decreasing production of $O_2^{\bullet-}$ and HO[•] [10].

Desferrioxamine can be used to inhibit lipid peroxidation. In systems containing metmyoglobin and methemoglobin, lipid peroxidation was inhibited almost completely by the addition of 10 μ M DFO. Interestingly, another chelator, EDTA, was used and had very little effect. The results are pictured in the graph below, taken from [17].



Figure 2- The effect of DFO and EDTA concentration on membranal lipid peroxidation by H_2O_2 -activated metmyglobin/methemoglobin. DFA drastically reduces peroxidation by this system.

Nitroxide Radical

Although desferrioxamine is an inhibitor of iron dependent free radical reactions, its prolonged use is far from benign. Actually, DFO can be oxidized by horseradish peroxidase [11], neutrophils [12], or other enzymes, and the resulting nitroxide radical



can initiate radical chemistry in a system.

Figure 3- The nitroxide radical produced from the reaction of DFO with HRP and H_2O_2 . The resolution of the resulting EPR spectrum is too poor to determine which hydroxamic acid is attacked. Adapted from [11].

The nitroxide radical is only formed at relatively high concentrations of DFO, greater than 1 mM. This means that DFO should not be used in biological systems with peroxidases present [11]. Neutrophils contain myeloperoxidase, which oxidizes DFO and causes problems with the removal of radicals from infection sites [12]. The nitroxide radical may also affect the activity of other enzymes, such as Ca²⁺-ATPase. This enzyme is found in the sarcoplasmic reticulum, and can act in the same manner as a peroxidase. Metabolites, such as desferrioxamine nitroxide radical, inhibit the enzyme by interrupting the Fenton reaction [13].

Detection of DFO Radical

Once desferrioxamine has been oxidized by a peroxidase, the EPR spectrum is obtainable, since the radical is rather stable. A sample spectrum, adapted from [11] is pictured below:



Figure 4- The one-electron oxidation of DFO by HRP. The reaction mixture, A, contained 0.1 mM Desferal, 0.1 mg/mL HRP, and 50 μ M H₂O₂ in a 100 mM phosphate buffer at pH 7.4. B-D were control experiments, each having 1 essential reactant removed. The gvalue is 2.0065, with $a^{\rm N} = 7.85$ G and $a^{\rm H} = 6.35$ G. Vladimirov *et al.* discovered a fantastic method for determining the free iron concentration in tissue homogenates. They used both DFO and 1,10-phenanthroline to chelate iron in both the (II) and (III) states. Then, a combination of EPR and UV/Vis techniques measured the concentration of free iron in rat liver [17].



Figure 5- Absorption spectra (A) and EPR signals (B) of solutions containing 50 μ M Fe(II) and 50 μ M Fe(III) in the presence of desferrioxamine (1), 1,10-phenanthroline (2), or a desferrioxamine-phenanthroline mixture (3). In A.3, an additive effect is observed, whereas in B.3 there is destructive interference. The EPR signal in B is centered at g = 4.3, indicating the presence of Fe(III). Adapted from [17].

Ferrioxamine absorbs at 430 nm ($\varepsilon = 2.48 \text{ mM}^{-1} \text{ cm}^{-1}$) [14], while desferrioxamine is a colorless crystal, so alternate methods are used to detect it. Other methods for detection of DFO include HPLC and FAB-MS. Reverse phase HPLC has been used to detect the amount of DFO available iron in rabbit kidneys. The kidneys were processed, with ferrioxamine and desferrioxamine eluting at different times. This experiment was done to check the total amount of iron which could be chelated by DFO, thus depriving other harmful enzymes, such as bleomycin, which uses iron to degrade DNA, of using iron to harm cells [15]. Finally, FAB-MS was used to test the urine of DFO-treated humans looking for the signature m/z of 613 [16]. This is probably one of the easiest methods of detection.

Conclusion

In summation, desferrioxamine is extremely effective at sponging up excess iron in biological systems. This is due to its high stability constant, as well as properties that make it friendly to most cells. In high concentrations however, a nitroxide radical may be formed, causing damage to the local environment. There are many methods used to detect desferrioxamine, including EPR and UV/Vis. These methods can also be used to discover interesting features of the cell, such as the iron content.

References

- [1] Schwarzenbach G, Schwarzenbach K. (1963) Properties of desferrioxamine. *Helv. Chim. Acta.*, 1390-1400.
- [2] Goodwin JF, Whitten CF. (1965) Desferrioxamine analysis. *Nature* 205:281-283.
- [3] Raymond KN, Muller G, Matzanke BF. (1984) Topics in Current Chemistry (Boschke, FL, Ed.) Vol 123, pp. 49-102, Springer-Verlag, Berlin.
- [4] Keberle, H. (1964) The biochemistry of desferrioxamine and its relation to iron metabolism. *Annals N.Y. Acad. Sci.* **119:**758-765.
- [5] Goldstein S, Czapski G. (1990) A reinvestigation of the reaction of desferrioxamine with superoxide radicals, a pulse radiolysis study. *Free Rad. Res. Comm.* 11:231-240.
- [6] K. Quirolo. <u>www.thassalemia.com/transfusion/chelation.shtml</u>. Children's Hospital of Oakland.
- [7] Bel A, Martinod E, Menasche P. (1996) Cardioprotective Effect of Desferrioxamine. *Acta. Haem.* **95:**63-65.
- [8] Roza AM Slakey DP Adams, MB *et al.* (1994) Hydroxylethyl starch deferrioxamine, a novel iron chelator, delays diabetes in BB rats. *J. Clin. Med.* 123: 556-560.
- [9] Lefebvre V, Buc-Calderon P. (1995) Desferal prevents against cell lysis induced by hydrogen peroxide to hypoxic hepatocytes: a role for free iron in hypoxiamediated cellular injury. *Chem.-Bio. Inter.* **94:**37-48.
- [10] Bhuyan KC, Bhuyan DK, Chiu W, Malik S, Fridovich I. (1991) Desferal-Mn(III) in the therapy of diquat-induced cataract in rabbit. *Arch. Biochem. Biophys.* 288:525-532.
- [11] Morehouse KM, Flitter WD, Mason RP. (1987) The enzymatic oxidation of desferal to a nitroxide free radical. *FEBS Lett.* **222**:246-250.
- [12] Soriani M, Mazzuca S, Quaresima V, Minetti M. (1993) Oxidation of desferrioxamine to nitroxide free radical by activated human neutrophils. *Free Radic. Biol. & Med.* 14:589-599.
- [13] Kiyose M, Lee C, Okabe E. (1999) Inhibition of skeletal sarcoplasmic reticulum Ca²⁺-ATPase activity by deferoxamine nitroxide free radical. *Chem. Res. Toxicol.* 12:137-143.

- [14] Miller DM, Spear NH, Aust SD. (1992) Effects of Deferrioxamine on ironcatalyzed lipid peroxidation. *Arch Biochem. and Biophys.* **295**:240-246.
- [15] Gower J, Healing G, Green C. (1989) Measurement by HPLC of desferrioxamine available iron in rabbit kidneys to assess the effect of ischaemia on the distribution of iron within the total pool. *Free Rad. Res. Comm.* **5**:291-299.
- [16] Lehmann WD, Heinrich HC. (1990) Ferrioxamine and its hexadentate iron chelating metabolites in human post-desferal urine studied by HPLC and FAB-MS. Anal. Biochem. 184:219-227.
- [17] Yegorov DY Kozlov AV Azizova OA Vladimirov YA. (1993) Simultaneous determination of Fe(III) and Fe(II) in water solutions and tissue homogenates using Desferal and 1,10-phenanthroline. *Free Radic. Biol. and Med.* 15:565-574.
- [18] Martell AE, Smith RM, Motekitis RJ. (1995) NIST critical stability constants of metal complexes database. Version 1.0.