

This student paper was written as an assignment in the graduate course

Free Radicals in Biology and Medicine

(77:222, Spring 2001)

offered by the

Free Radical and Radiation Biology Program

B-180 Med Labs

The University of Iowa

Iowa City, IA 52242-1181

Spring 2001 Term

Instructors:

GARRY R. BUETTNER, Ph.D.

LARRY W. OBERLEY, Ph.D.

with guest lectures from:

Drs. Freya Q. Schafer, Douglas R. Spitz, and Frederick E. Domann

The Fine Print:

Because this is a paper written by a beginning student as an assignment, there are no guarantees that everything is absolutely correct and accurate.

In view of the possibility of human error or changes in our knowledge due to continued research, neither the author nor The University of Iowa nor any other party who has been involved in the preparation or publication of this work warrants that the information contained herein is in every respect accurate or complete, and they are not responsible for any errors or omissions or for the results obtained from the use of such information. Readers are encouraged to confirm the information contained herein with other sources.

All material contained in this paper is copyright of the author, or the owner of the source that the material was taken from. This work is not intended as a threat to the ownership of said copyrights.

Hydroxyl Radical

by

Jianfang Hu

B-180 Medical Laboratories

Free Radical and Radiation Biology Program

The University of Iowa
Iowa City, IA 52242-1181

For 77:222, Spring 2001
Feb. 8th. 2001

Abbreviation:

DMPO: 5, 5-dimethylpyrroline-N-oxide;

UV: ultraviolet;

EPR: electron paramagnetic resonance;

DEPMPO: 5,5-dimethyl-pyrroline-N-oxide and 5-diethoxyphosphoryl-5-methyl-pyrroline-N-oxide

Contents

I.	Abstract	2
II.	Introduction	3
III.	Generation	3
	1. Radiolysis of water	3
	2. Fenton reaction	4
	3. Photolysis	4
	4. Ozonation	5
	5. Sonolysis	5
IV.	Detection	5
V.	Reactions	6
VI.	Hydroxyl radical in cell function	7
VII.	Summary	8
VII.	Reference	9

I. Abstract:

Hydroxyl radical is a highly oxidizing free radical. It is widely produced in many biological processes, and causes various oxidative damage in organisms. Hydroxyl radical is generated from one-electron reduction of hydrogen peroxide by metal ions and /or organic molecules *in vivo*, therefore it is highly electrophilic in nature. Understanding the properties of hydroxyl radical can help develop methods to reduce oxidative damage. The generation, detection, and reactions of this radical are reviewed in this paper.

Introduction

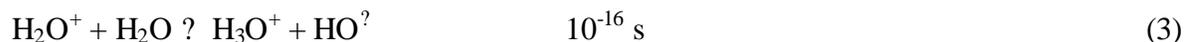
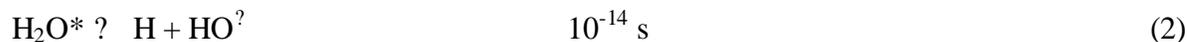
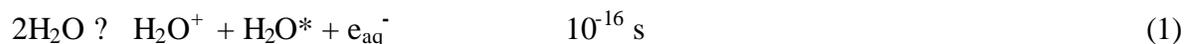
Hydroxyl radical is a highly reactive and toxic radical with one unpaired electron. It is formed through one-electron reduction of hydrogen peroxide [1]. It is highly electrophilic in nature [2]. Thus it is a powerful oxidizing agent. It is formed in living cells during physiological processes or after environment stimulation. Since it is a highly reactive radical, once it is formed, it will react with various biomolecules. This then causes different biological damage, which may eventually cause many disease states. Aging, heart disease, Parkinson's disease, and rheumatoid arthritis are thought to be partial outcomes of hydroxyl radical damage. In order to make progress in our understanding of these areas, it is necessary to study the chemical properties of this radical, such as the generation, detection, and reactivity.

Generation

There are a number of ways to generate hydroxyl radical, some of them are used *in vitro* as research tools. Water radiolysis, Fenton-type reaction, photolysis, sonolysis, and ozonation are briefly introduced below.

Radiolysis of water

When water is exposed to ionizing radiation, such as x-ray and γ -ray, hydroxyl radical and other radicals are produced [4]. Cell killing caused by ionizing radiation is mostly due to the hydroxyl radicals produced by water radiolysis [3]. The reactions are listed below:

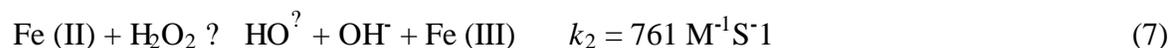


H_2O^* is an excited water molecule, and e_{aq}^- is a hydrated electron. When N_2O is present, the reaction is [3]:



Fenton reaction

Classical Fenton reaction is referred as superoxide interaction with hydrogen peroxide to produce hydroxyl radical. This reaction is thermodynamically favored, ($\Delta G = -18.2$ kcal/mol) [22] but the activation energy prevents it from happening spontaneously. The rate constant can be increased dramatically in the presence of catalysts, such as iron or copper. The overall reaction was first proposed by Haber and Weiss in 1934 [5], therefore it is also named as Haber-Weiss reaction.



In 1998, hydroxyl radical was detected when H_2O_2 reacted with nitric oxide in the absence of transition metals in Vass' lab [6]. Recently, M. Chevion found tetrachlorohydroquinon (TCHO) and hydrogen peroxide complex also had the ability to generate hydroxyl radical in a metal-independent way. The mechanism is unclear [7].

Photolysis

Photolysis is another important way to produce hydroxyl radical. UVB (280-320 nm) is responsible for this reaction. Under UV light exposure, intracellular H_2O_2 production is increased, which in turn forms in the presence of transition metals in a Fenton-type reaction [8].



UV can directly split H_2O_2 into two HO^\cdot [23]:



Ozonation

Ozone is a slowly reacting oxidant. It can transform into other more reactive oxygen species in aqueous solution, such as hydroxyl radical, and carbon centered radicals [9]. Hoigen and Bader first reported that hydroxyl radicals were oxidizing intermediates in ozonation of water. [10]. The reaction equation is:



Sonolysis

Water vapor at very high temperatures and pressures can be decomposed to HO^\cdot . These temperatures and pressures are achieved in the gas bubbles by ultrasound. The decomposition reaction is [4]:



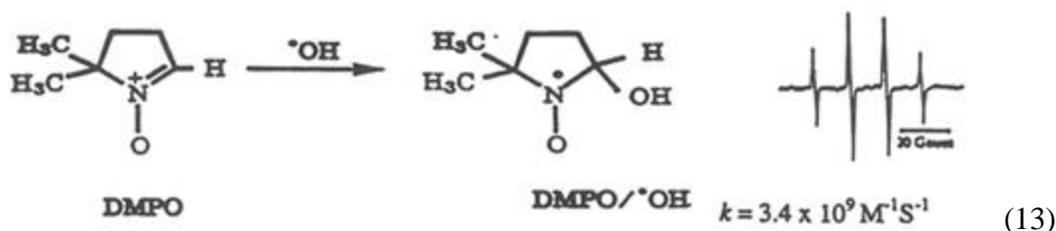
In summary, hydroxyl radical is produced in various physiological and pathological processes, and it can also be produced after certain chemical and physical stimulations, such as sunlight, smoke. The basic reactions are radiolysis and Fenton-type reaction.

Detection

Hydroxyl radical is extremely reactive. Its half-life is approximately 10^{-9} second [11]. It cannot be detected directly. Electron paramagnetic resonance (EPR) spin-trapping and high performance liquid chromatography electrochemical detection (HPLC-EC) are two widely used detection methods.

EPR

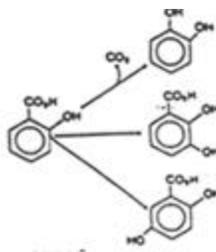
Free radicals have one or more unpaired electrons. This is the reason for its paramagnetic property. In order to detect the hydroxyl radical, spin trap agents should react with it to make more stable nitroxide radicals, these relatively stable radicals can be observed by EPR. Some commonly used spin trapping agents are DMPO [12], DEPMPO [13], salicylate [15]. Here is the reaction between DMPO and HO[·]:



“The low stability of Hydroxyl radical-derived nitroxides is a limiting factor for directly spin trapping of HO[·] in biological systems.” Addition of DMSO can partly solve this problem because HO[·] oxidizes DMSO to methyl radical. The nitroxide derivatives of methyl radical are more stable than other spin-trapped nitroxides [14]. Equation (14) shows the chemistry.

**HPLC-EC**

Hydroxyl radical reacts with a number of molecules by hydroxylation. These hydroxylation products can be separated in HPLC, then detected by electrochemical detection, or fluorescence. Salicylate, tyrosine, and Phenylalanine [16] are some commonly used agents. The figure showed below is hydroxyl radical reacting with salicylate. The rate constant is $1.2 \times 10^{10} \text{ M}^{-1}\text{s}^{-1}$ [17]:



(11%) Catechol, (49%) 2,3-Dihydroxybenzoate, (40%) 2,5-Dihydroxybenzoate

Figure 1: Chemical trapping of HO[·] by salicylate. [Adapted from 17].

Reactions

Hydroxyl radical rapidly reacts with other molecules in three main types reactions. All of these reactions are very fast, the rate constants are $10^9 \text{ M}^{-1}\text{s}^{-1}$ magnitude [3]:

1). Hydrogen atom abstraction:

For example, when hydroxyl radical reacts with ethanol, it abstracts a hydrogen atom from ethanol to form water [3]:



2). Electron- transfer:

Sometimes, hydroxyl radical transfers its electron to organic/inorganic compounds to make OH⁻:



3) Addition reaction:

Hydroxyl radical can add to ring structures with unsaturated bonds. .

(18)

II. Radical-radical reactions

Hydroxyl radical reacts with other radicals at nearly diffusing limiting rates because these reactions require no activation energy [4]. Some of these reactions are listed below:



Hydroxyl radical in cell function

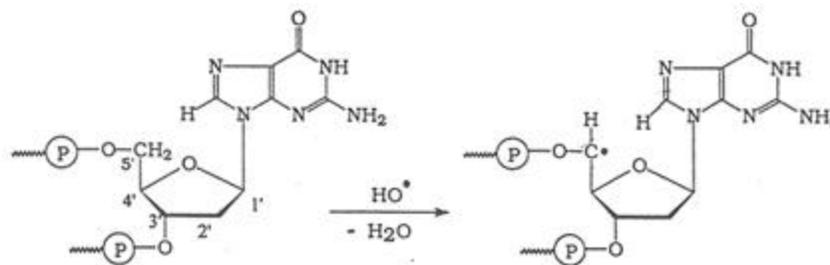
Activated neutrophils release hypochlorous acid (HOCl) and $\text{O}_2^{\cdot-}$, which in turn forms hydroxyl radical. This is due to the cytotoxic effects of the neutrophils [18]. The reaction is:



NADH, GSH and H_2O_2 are involved in respiration, it is reported that GSH and NADH can react with H_2O_2 to produce hydroxyl radical, which has considerable biological significance [19].



Hydroxyl radical can quickly reacts with all kinds of cell components, such as protein, lipid, and DNA/RNA. It reacts with amino acid by decarboxylation [20]. Equation 22 shows the chemistry. It reacts with DNA by abstracting a hydrogen atom from the deoxyribose sugars along the DNA backbone, and cut down the DNA backbone. Equation 23 shows the reaction [24].



(23)

Summary

In short, hydroxyl radical is generated in various conditions including physiological and pathological processes. It is powerful oxidant, it reacts with most molecules in a nearly diffusion

limiting rate. Though direct detection of this radical is impossible, a number of indirect methods have been developed. Further research in its chemical and biological properties can lead to better understand of many pathological processes.

References

1. Koppenaol WH, and Liebman J F. (1984) *The journal of physical chemistry*. **88**: 99-101.
2. Imamura A., and Hirao K. (1979) A molecular orbital approach to the eletrophilicity of H and OH radical. *Bulletin of the cheical society of Japan*. **52** (2): 287-292.
3. Baxton GV, Greenstock CL, Helman WP, Ross AB. (1988) Critical review of rate constants for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals (OH[•]/O) in aqueous solution. *J. Phys. Chem. Ref. Data*. **17**: 513-886.
4. Nappi AJ, Vass E. (1998) Hydroxyl radical formation resulting from the interaction of nitric oxide and hydrogen peroxide. *Biochimica Biophys Acta*. **1380** (1): 55-63.
- 5 Roots R, Okada S. (1975) Estimation of life times and diffusion distances of radicals involved in X-ray induced DNA strand breaks or killing of mammalian cells. *Radiat. Res*. **64**: 306-320.
6. Haber F and Weiss J (1934) The actalytic decomposition of hydrogen peroxide b iron salts. Proceeding of the Royal of Lon: *Methe. And Physical Society*. **147**: 332-351.
- 7..Zhu BZ, Kitrossky N, and Chevion M. (2000) Evidence for production of hydroxyl radical by pentachlorophenol metabolites and hydrogen peroxide: A metal-independent organic Fenton reaction. *Biochem. Bophys. Res.Communications*. **270** (3): 942-946.
8. Masaki H, astumi T, Sakurai H. (1995) Deteection of hydrogen peroxide and hydroxyl radicals in murine skin fibroblasts under UVB irradiation. *Biochem Biophys Res Comm*. **206**: 474-479.
- 9Ueno I., Hoshino M, T. Miura T, Shinrki N. (1998) Ozone exposure generates free radicals in the blood samples in vitro: detection by the ESR spin-trapping technique. *Free Radical Research*. **29**(2): 127-135.
10. Hoigen J, Bader H (1975) Ozonation of water: role of hydroxyl radicals as oxidizing intermediates. *Science*. **190**:782-784.
11. Bveris A. (1998) Biochemistry of radicals: from electron to tissues. *Medicine*. **58**(4): 350-356.

12. Buettner. GR. (1978) Considerations in the spin trapping of superoxide and hydroxyl radical in aqueous systems using DMPO. *Biochem.Bioph Res.Comm.* **83**: 69-74.
13. Timmins GS, Liu KJ, Bechara EJ, Kotake Y, Swartz HM. (1999) Trapping of free radicals with direct in vivo EPR detection: a comparison of 5,5-dimethyl-pyrroline-N-oxide and 5-diethoxyphosphoryl-5-methyl-pyrroline-N-oxide as spin traps for $\cdot\text{OH}$ and $\text{SO}_4^{\cdot-}$. *Free radical Biology & Medicine.* **27**(3-4): 329-333.
14. Stoyanovsky DA, Melenikov Z, Cederbaum AI. (1999) ESR and HPLC-EC analysis of the interaction of hydroxyl radical with DMSO: rapid reduction and quantification of POBN and PBN nitroxides. *Analytical Chemistry.* **71**(3): 715-721.
15. Powell SR. (1994) Commentary salicylate trapping of $\cdot\text{OH}$ as a tool for studying post-ischemic oxidative injury in the isolated rat heart. *Free Rad. Res.* **21**(6): 355-370.
16. Maskos A, Rush JD, Koppenol WH. (1992) The hydroxylation of phenylalanine and tyrosine: a comparison with salicylate and tryptophan. *Arch. of Biochem. Biophy.* **296**(2): 521-529.
17. Halliwell B, Gutteridge JM. (1989) Free radicals in Biology and Medicine. *Clarendon. Press.* Oxford.
18. Umeda T, Hara T, Hayashida M and Nijima T. (1985) Role of hydroxyl radical in neutrophil-mediated cytotoxicity. *Cel. Mol. Biol.* **31**(3): 229-233.
19. Florence. TM (1984) The production of hydroxyl radical from the reaction between hydrogen peroxide and NADH. *J. Inorganic Biochem.* **28**:33-37.
20. Steffen. L. K., Glass R.S., Sabahi M., Wilson G. S., Schoneich C., Mahling S., and Asmus K. D. (1991) HO^{\cdot} radical induced decarboxylation of amino acid. Decarboxylation vs bond formation in radical intermediates. *J. Ame. Chem. Soci.* **113**(6): 2141-2145.
21. Tullius TD, Dombroski BA, Churchill MEA, and Kam L. (1987) Hydroxyl radical footprinting: a high-resolution method for mapping protein-DNA contacts. *Methods in enzymology.* **15**. 537-539.
22. Buettner GR, Oberley LW. (2001) *Course notes in free radical in biology and medicine.* Chapter **IV**:12.
23. Buettner GR, Oberley LW. (2001) *Course notes in free radical in biology and medicine.* Chapter **V**:2.
24. Buettner GR, Oberley LW. (2001) *Course notes in free radical in biology and medicine.* Chapter **XII**: 11.